

Wave Life Sciences Corporate Presentation

March 2, 2020



Forward-looking statements

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Building a leading genetic medicines company

PRISM

INNOVATIVE PLATFORM

- Stereopure oligonucleotides
- Backbone modifications
- Allele-selectivity
- Novel modalities (ADAR)
- Foundational stereochemistry IP



FOUNDATION OF CNS PROGRAMS

- Huntington's disease
- ALS / FTD
- Ataxias
- Parkinson's
- Alzheimer's



CLINICAL DEVELOPMENT EXPERTISE

- Multiple global clinical trials ongoing across eight countries
- Innovative trial designs



MANUFACTURING

 Established internal manufacturing capabilities to produce oligonucleotides at scale

Innovative pipeline led by CNS programs

THERAPEUTIC AREA	TARGET	DISCOVERY	PRECLINICAL	CLINICAL	ESTIMATED U.S. PREVALENCE*	PARTNER
CNS						
	WVE-120101 mHTT SNP1		Phase 1b/	2a and OLE	~10,000 / ~35,000	Takeda 50:50 option
Huntington's disease	WVE-120102 mHTT SNP2		Phase 1b/	2a and OLE	~10,000 / ~35,000	Takeda 50:50 option
	mHTT SNP3				~8,000 / ~30,000	Takeda 50:50 option
ALS and FTD	C9orf72				~1,800 (ALS) ~7,000 (FTD)	Takeda 50:50 option
Spinocerebellar ataxia 3	ATXN3				~4,500	Takeda 50:50 option
CNS diseases	Multiple ⁺					Takeda milestones & royalties
OPHTHALMOLOGY						
Retinal diseases	USH2A and RhoP23H					100% global
HEPATIC						
Metabolic liver diseases	Multiple					Pfizer milestones & royalties
OTHER						
ADAR RNA-editing	Multiple					100% global



*Estimates of U.S. prevalence and addressable population by target based on publicly available data and are approximate; for Huntington's disease, numbers approximate manifest and pre-manifest populations, respectively.

[†]During a four-year term, Wave and Takeda may collaborate on up to six preclinical targets at any one time.

ALS: Amyotrophic lateral sclerosis; FTD: Frontotemporal dementia; CNS: Central nervous system; OLE: Open-label extension

LIFE SCIENCES

HD portfolio Huntington's Disease

Huntington's disease: a hereditary, fatal disorder

- Autosomal dominant disease, characterized by cognitive decline, psychiatric illness and chorea; fatal
- No approved disease-modifying therapies
- Expanded CAG triplet repeat in HTT gene results in production of mutant huntingtin protein (mHTT); accumulation of mHTT causes progressive loss of neurons in the brain
- Wild-type (healthy) HTT protein critical for neuronal function; evidence suggests wild-type HTT loss of function plays a role in Huntington's disease
- 30,000 people with Huntington's disease in the US; another 200,000 at risk of developing the condition





Sources: Auerbach W, et al. Hum Mol Genet. 2001;10:2515-2523. Dragatsis I, et al. Nat Genet. 2000;26:300-306. Leavitt BR, et al. J Neuroschem. 2006;96:1121-1129. Nasir J, et al. Cell. 1995;81:811-823. Reiner A, et al. J Neurosci. 2001;21:7608-7619. White JK, et al. Nat Genet. 1997;17:404-410. Zeitlin S, et al. Nat Genet. 1995;11:155-163. Carroll JB, et al. Mol Ther. 2011;19:2178-2185. HDSA 'What is Huntington's disease? https://hdsa.org/what-is-hd/overview-of-huntingtons-disease/ Accessed: 11/2/18.; Becanovic, K., et al., Nat Neurosci, 2015. 18(6): p. 807-16. Van Raamsdonk, J.M., et al., Hum Mol Genet, 2005. 14(10): p. 1379-92.; Van Raamsdonk, J.M., et al., BMC Neurosci, 2006. 7: p. 80.

Importance of wild-type huntingtin (wtHTT) in HD

Huntington's disease (HD) may be caused by a dominant gain of function in mutant HTT and a loss of function of wtHTT protein

- Evidence suggests wild-type or healthy HTT is neuroprotective in an adult brain
 - Transport of key neurotrophic factors such as brain-derived neurotrophic factor (BDNF) are regulated by wtHTT levels
- Relative proportion of wild-type to mutant protein is critical
 - Increased amount of wild-type protein relative to mutant HTT may result in slower disease progression (measured by age-at-onset)
 - Patients with lack of wild-type have significantly more severe disease (measured by disease progression after symptom onset)







Sources: Van Raamsdonk, J.M., et al., Hum Mol Genet, 2005; Van Raamsdonk, J.M., et al., BMC Neurosci, 2006; Becanovic, K., et al., Nat Neurosci, 2015; Saudou, F. and S. Humbert, The Biology of Huntingtin. Neuron, 2016; Gauthier, L.R., et al., Cell, 2004; Caviston, J.P. and E.L. Holzbaur, Trends Cell Biol, 2009; Ho, L.W., et al., J Med Genet, 2001, Zuccato et al., Science 2001; Zuccato et al., Brain Pathol 2007; Marullo et al. Genome Biol 2010; Squitieri et. al, Brain 2003

Increasing evidence on the importance of wtHTT in HD pathogenesis, CNS and systemic health

Recent publications on wtHTT LoF as a likely driver of HD pathogenesis



wtHTT in HD highlighted at CHDI 15th Annual HD Therapeutics Conference:

HTT LOWERING: EXPLORING DISTRIBUTION, TIMING, AND SAFETY (LOSS OF FUNCTION)

Key points discussed at meeting:

- wtHTT has numerous critical functions throughout life (e.g., intracellular trafficking, cell-cell adhesion, BDNF transport)
- Near elimination of mouse wtHtt detrimental regardless of when suppression begins
- Specific brain regions, e.g., STN, may be particularly vulnerable to wtHTT lowering
- Mouse Htt lowering can lead to thalamic, hepatic, pancreatic toxicity
- HTT LoF mutations highly constrained in human population, suggesting selection against LoF mutations

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LoF: Loss of function; wtHTT: wild-type huntingtin; HD: Huntington's disease; STN: subthalamic nucleus

Wild-type HTT in the cortex appears critical for striatal health



Status of the presynaptic compartment determines the integrity of the network



Presented by Dr. Frederic Saudou at Wave's Analyst and Investor Research Day on October 7, 2019 Virlogeux et al., Cell Reports 2018

Wave approach: novel, allele-selective silencing

Aims to lower mHTT transcript while leaving healthy wild-type HTT relatively intact

- Utilize association between single nucleotide polymorphisms (SNPs) and genetic mutations to specifically target errors in genetic disorders, including Huntington's disease (HD)
- Potential to provide treatment for up to 80% of HD population





Allele-selectivity possible by targeting SNPs associated with expanded long CAG repeat in HTT gene



Source: Kay, et al. Personalized gene silencing therapeutics for Huntington disease. Clin Genet. 2014;86:29-36.

Selective reduction of mHTT mRNA & protein



Reporter Cell Line*



Source: Meena, Zboray L, Svrzikapa N, et al. Selectivity and biodistribution of WVE-120101, a potential antisense oligonucleotide therapy for the treatment of Huntington's disease. Paper presented at: 69th Annual Meeting of the American Academy of Neurology; April 28, 2017; Boston, MA.

^{*}These results were replicated in a patient-derived cell line

Demonstrated delivery to brain tissue

• WVE-120101 and WVE-120102 distribution in cynomolgus non-human primate brain following intrathecal bolus injection



treatment of Huntington's disease. Paper presented at: 69th Annual Meeting of the American Academy of Neurology; April 28, 2017; Boston, MA.

PRECISION-HD clinical trial design

Two parallel, multicenter, double-blind, randomized, placebo-controlled Phase 1b/2a clinical trials for WVE-120101 and WVE-120102



Initiated February 2020



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Multidose Cohorts N = 12 per cohort

- PRECISION-HD2 data from 32 mg cohort expected in 2H 2020
- PRECISION-HD1 topline data, including 32 mg cohort, expected in 2H 2020

PRECISION-HD2 topline results

Clinical trial ongoing

Doses	Safety	Biomarker Effects	
		mHTT	wtHTT
• WVE-120102 2–16 mg (pooled)	 Generally safe and well tolerated 	 Reduction in mHTT compared to placebo (-12.4%¹, p<0.05²) Analysis across groups suggests dose response at highest doses (p=0.03)³ 	 No change in tHTT compared to placebo Ongoing evaluation
 32 mg cohort initiated Assessing the potential for higher dose cohorts 	 Safety profile supports addition of higher dose cohorts 	 Potential for greater mHTT reduction at higher doses 	 Larger reductions of mHTT expected to result in discernible impact on tHTT



Topline results announced December 30. 2019; mHTT: mutant huntingtin wtHTT: wild-type HTT tHTT: total HTT ¹ Hodges-Lehmann non-parametric shift estimates of the difference between treatment and placebo; ² Wilcoxon-Mann-Whitney non-parametric significance test; ³ Multiple Contrast Test (MCT)

Three allele-selective HD programs

Potential to address ~80% of HD patient population



% Huntington's Disease Patient Population with SNP

Intend to explore efficacy in early manifest and pre-manifest HD patient populations



SNP3 program approaching clinical development

Wave SNP3

Compound-2

-2 -1

wtHTT

mRNA

-3



Wave SNP3

Compound-1

Ω

Log₁₀ (µM compound concentration)

Knockdown persists for 12 weeks in BACHD mouse model



Cortex



Relative HTT mRNA expression

0.7-

0.5-

0.4-0.3-

0.2-

01

0.0

-3 -2

Pan-silencing

active comparator

-1 0

2 -3 -2 -1

Data presented at CHDI Foundation's 15th Annual HD Therapeutics Conference Feb 24-27, 2020; See poster for full dataset. [Figure on right] Statistics: All oligo treatment groups statistically significantly different from PBS; One-way ANOVA ****, P≤0.0001. SNP3 Compound-1 and Compound-2 significantly different from pansilencing active comparator at 8, 12 weeks ***, P<0.005; **P=0.001."</p>

Clinical development expected to initiate in 2H 2020

mHTT

mRNA

16

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PRISM Platform



Enables Wave to target genetically defined diseases with stereopure oligonucleotides across multiple therapeutic modalities



OPTIMIZE

A deep understanding of how the interplay among oligonucleotide sequence, chemistry, and backbone stereochemistry impacts key pharmacological properties

Through iterative analysis of *in vitro* and *in vivo* outcomes and artificial intelligence-driven predictive modeling, Wave continues to define design principles that are deployed across programs to rapidly develop and manufacture clinical candidates that meet pre-defined product profiles



PRISM enables optimal placement of backbone **PRISM** stereochemistry

Crystal structure confirms phosphate-binding pocket of RNase H binds 3'-SSR-5' motif in stereopure oligonucleotide – supports design strategy for Wave oligonucleotides



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Importance of controlling stereochemistry

Stereochemical diversity



Exponential diversity arises from uncontrolled stereochemistry





PRISM platform enables rational drug design



Source: Iwamoto N, et al. Control of phosphorothioate stereochemistry substantially increases the efficacy of antisense oligonucleotides. Nat Biotechnol. 2017;35:845-851.

Optimizing potency and durability across multiple tissues





Data represented in this slide from *in vivo* studies. CNS: PBS = phosphate buffered saline; Ctx = cortex; Str = striatum; Cb = cerebellum; Hp = hippocampus; SC = spinal cord. ICV = intracerebral; IVT = intravitreal; IV = intravenous; SC= subcutaneous.

LIFE SCIENCES

C9orf72 program

Amyotrophic Lateral Sclerosis (ALS) Frontotemporal Dementia (FTD)

C9orf72: a critical genetic risk factor

- C9orf72 gene provides instructions for making protein found in various tissues, with abundance in nerve cells in the cerebral cortex and motor neurons
- C9orf72 genetic mutations are the strongest genetic risk factor found to date for the more common, non-inherited (sporadic) forms of Amyotrophic Lateral Sclerosis (ALS) and Frontotemporal Dementia (FTD); GGGGCC repeat drives the formation and accumulation of dipeptide repeat proteins that accumulate in brain tissue
- First pathogenic mechanism identified to be a genetic link between familial (inherited) ALS and FTD
- Most common mutation identified associated with familial ALS and FTD
- Availability of dipeptide biomarker in CSF has potential to accelerate drug development



Amyotrophic lateral sclerosis

- Fatal neurodegenerative disease characterized by the progressive degeneration of motor neurons in the brain and spinal cord
- Affects approximately 15,000-20,000 people in the US with a median survival of three years
- C9orf72 is present in approximately 40% of familial ALS and 8-10% of sporadic ALS; currently the most common demonstrated mutation related to ALS, far more so than SOD1 or TDP-43
- Pathogenic transcripts of the C9orf72 gene contain hundreds to thousands of hexanucleotide repeats compared to 2-23 in wild-type transcripts; dominant trait with high penetrance





Frontotemporal dementia

- Progressive neuronal atrophy with loss in the frontal and temporal cortices characterized by personality and behavioral changes, as well as gradual impairment of language skills
- Affects approximately 55,000 people in the US
- Second most common form of early-onset dementia after Alzheimer's disease in people under the age of 65
- Up to 50% of FTD patients have a family history of dementia, many inheriting FTD as an autosomal dominant trait with high penetrance
- Pathogenic transcripts of the C9orf72 gene contain hundreds to thousands of hexanucleotide repeats compared to 2-23 in wild-type transcripts





Sources: Stevens M, et al. Familial aggregation in frontotemporal dementia. *Neurology.* 1998;50:1541-1545. Majounie E, et al. Frequency of the C9orf72 hexanucleotide repeat expansion in patients with amyotrophic lateral sclerosis and frontotemporal dementia: a cross-sectional study. *Lancet Neurol.* 2012;11:323-330.

C9orf72 program: Selective silencing *in vivo* of 4 expanded C9orf72 repeat transcripts

- C9orf72 genetic mutations are the most common cause of familial Amyotrophic Lateral Sclerosis (ALS) and Frontotemporal Dementia (FTD) and are the strongest genetic risk factor found to date for the more common, non-inherited (sporadic) forms of ALS and FTD; Hexanucleotide repeat drives the formation and accumulation of dipeptide repeat proteins that accumulate in brain tissue
- Wave's approach: Selectively silence the repeat containing transcript while minimizing the impact on C9orf72 protein



Experimental description: 2 x 50 ug on day 1 and day 8; mRNA Samples were analyzed using quantitative PCR (Taqman assay), Protein samples were measured by Western Blot. Dipeptide repeat proteins were measured by Poly-GP MSD assay.

Neuro C9orf72

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Ophthalmology

Stereopure oligonucleotides for inherited retinal diseases (IRDs)

Wave ophthalmology opportunity

- Oligonucleotides can be administered by intravitreal (IVT) injection; targeting twice per year dosing
- Stereopure oligonucleotides open novel strategies in both dominant and recessive IRDs; potential for potent and durable effect with low immune response

Successful targeting of *MALAT1* is a surrogate for an ASO mechanism of action

- Widely expressed in many different cell types
- Only expressed in the nucleus



Intravitreal injection



Sources: Daiger S, et al. *Clin Genet*. 2013;84:132-141. Wong CH, et al. *Biostatistics*. 2018; DOI: 10.1093/biostatistics/kxx069. Athanasiou D, et al. *Prog Retin Eye Res*. 2018;62:1–23. Daiger S, et al. *Cold Spring Harb Perspect Med*. 2015;5:a017129. Verbakel S, et al. *Prog Retin Eye Res*. 2018:66:157-186.; Short, B.G.; *Toxicology Pathology*, Jan 2008.

Stereopure compound induces potent and durable *MALAT1* knockdown in the eye





Mouse: Compound or PBS (1 x 50 mg IVT) was delivered to C57BL6 mice. Relative percentage of Malat1 RNA in the posterior of the eye (retina, choroid, sclera) to PBS-treated mice is shown at 12, 20 and 36 weeks post-single injection. Statistics: Compound-2 Malat1 levels are significantly different from NTC at 36 weeks ***, P<0.001; **** P<0.0001, respectively. PBS = phosphate buffered saline; NTC= chemistry matched non-targeting control; Compound-1 and Compound-2 are stereopure MALAT1-targeting antisense oligonucleotide. NHP: Oligonucleotide or PBS (1 x 450 µg IVT) was delivered to NHP. Relative percentage of *MALAT1* RNA in the retina to PBS-treated is shown at 1 week, 2 and 4 months, post-single injection. Compound-1 is a stereopure *MALAT1*-RNA-targeting antisense oligonucleotide.

Ophthalmology

Usher Syndrome Type 2A: a progressive vision loss disorder

- Autosomal recessive disease characterized by hearing loss at birth and progressive vision loss beginning in adolescence or adulthood
- Caused by mutations in USH2A gene (72 exons) that disrupt production of usherin protein in retina, leading to degeneration of the photoreceptors
- No approved disease-modifying therapies
- ~5,000 addressable patients in US



Oligonucleotides that promote USH2A exon 13 skipping may restore production of functional usherin protein



Sources: Boughman et al., 1983. J Chron Dis. 36:595-603; Seyedahmadi et al., 2004. Exp Eye Res. 79:167-173; Liu et al., 2007. Proc Natl Acad Sci USA 104:4413-4418.

Potent USH2A exon 13 skipping with stereopure compound in *vitro* and *ex vivo*



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Left: Compounds were added to Y79 cells under free-uptake conditions. Exon skipping was evaluated by Taqman assays. *USH2A* transcripts were normalized to *SRSF9*. Data are mean \pm s.d., n=2. Reference Compound: van Diepen *et al.* 2018. Antisense oligonucleotides for the treatment of eye disease. W02018055134A1. Compound-1 is a stereopure antisense oligonucleotide. Right: Whole NHP and human eyes were enucleated (n=4 and n=2, respectively) and compounds (1-20 µM) were added to extracted retinas under free-uptake conditions. Exon skipping was evaluated by 48 hrs later by Taqman assays on RNA. USH2A transcript levels were normalized to SRSF9. Data presented are mean \pm s.e.m.

Ophthalmology

Allele-selective reduction of SNP-containing allele for adRP associated with Rhodopsin P23H mutation

- **Retinitis pigmentosa (RP)**: group of rare, genetic eye disorders resulting in progressive photoreceptor cell death and gradual functional loss; currently no cure
- ~10% of US autosomal dominant RP cases are caused by the P23H mutation in the rhodopsin gene (RHO)
- Mutant P23H rhodopsin protein is thought to misfold and co-aggregate with wild-type rhodopsin, resulting in a gain-of-function or dominant negative effect in rod photoreceptor cells



Ferrari et al., Current Genomics. 2011;12:238-249.; Reporter assays on a Wave stereopure sequence as well as a sequence described in WO2016138353A1: ASO and luciferase reporter plasmids (wild-type and mutant rhodopsin) are transfected into Cos7 cells. 48-hours later, cells are harvested, and relative luminescence is measured.

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ADAR-mediated RNA editing

RNA-editing can be used for several therapeutic applications and supplement Wave's existing modalities

		Treatment Modality		
Strategy	Therapeutic Application	Silencing Splicing RM	A Editing	
Silence protein expression	Reduce levels of toxic mRNA/protein	\checkmark	\checkmark	
Alter mRNA splicing	Exon skipping/inclusion/ restore frame	\checkmark	\checkmark	
Fix nonsense mutations that cannot be splice-corrected	Restore protein expression		\checkmark	
Fix missense mutations that cannot be splice-corrected	Restore protein function		\checkmark	
lodify amino acid codons Alter protein function			\checkmark	
Remove upstream ORF	Increase protein expression	Edited RNA	 ✓ 	

Using PRISM to unlock ADAR-mediated RNA editing

Structure of ADAR deaminase domain bound to dsRNA substrate



- ADAR makes multiple contacts with oligonucleotide backbone, sugar and bases
- Using PRISM platform, rationally designed and screened oligonucleotides to optimize:
 - 2' sugar chemistry
 - Backbone chemistry and stereochemistry
 - Size and structure
 - Modified nucleobases

~1,000 RNA editing oligonucleotides tested over the last year to develop SAR for editing format



Structure adapted from Matthews et al., Nat Struct Mol Biol. (2016); SAR = structure-activity relationship; ADAR: Adenosine Deaminase Acting on RNA; dsRNA = double-stranded RNA

Wave's ADAR approach has several potential advantages over existing technologies



Single oligonucleotide through free uptake is sufficient for editing



RNA editing

RNA Editing with Endogenous ADAR Achieved Across Multiple Primary Human Cell Types



Technology Validated Across Multiple Sequences In Vitro

Editing in Primary Human Hepatocytes



• Editing achieved across several distinct RNA transcripts

In vivo editing data with fully modified stereopure oligonucleotides expected in 2020



Data presented at 1st International Conference on Base Editing - Enzymes and Applications (Deaminet 2020); See poster for full dataset

RNA editing

Wave three-year outlook

	EXECUTE	EXPAND	EVOLVE			
CNS	PRECISION-HD trials Phase 1b/2a results	Four clinical programs, including	SNP3 and C9orf72 clinical data			
	SNP3 and C9orf72 clinical development initiations	potential pivotal HD trials	Other ongoing clinical programs			
	Advance Takeda and V	Multiple potential CTA filings				
Eye	Advance USH2A and RhoP23H					
Liver Other	ADAR RNA-editing POC	ADAR RNA-editing platform	ADAR RNA-editing programs			
	<i>in vivo</i> data	development	New non-CNS targets			
	2020	2021	2022			
WAVE Chemistry innovations transferred to new programs						
LIFE SCIENCES POC: proof of concept						

Anticipated upcoming Wave milestones

CNS

- 2H 2020: PRECISION-HD2 data from 32 mg cohort in Huntington's disease
- 2H 2020: PRECISION-HD1 topline data, including 32 mg cohort, in Huntington's disease
- 2H 2020: Initiate clinical development of SNP3 program in Huntington's disease
- 2H 2020: Initiate clinical development of C9orf72 program in ALS and FTD

Ophthalmology

• 2020: Advance USH2A and RhoP23H programs

RNA-editing

• 2020: In vivo ADAR editing data



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Realizing the potential of genetic medicines

For more information:

Kate Rausch, Investor Relations krausch@wavelifesci.com 617.949.4827