



Wave Life Sciences  
R&D Day

September 28, 2023

**WAVE**<sup>®</sup>  
LIFE SCIENCES

# Forward-looking statements

This document contains forward-looking statements. All statements other than statements of historical facts contained in this document, including statements regarding possible or assumed future results of operations, preclinical and clinical studies, business strategies, research and development plans, collaborations and partnerships, regulatory activities and timing thereof, competitive position, potential growth opportunities, use of proceeds and the effects of competition are forward-looking statements. These statements involve known and unknown risks, uncertainties and other important factors that may cause the actual results, performance or achievements of Wave Life Sciences Ltd. (the “Company”) to be materially different from any future results, performance or achievements expressed or implied by the forward-looking statements. In some cases, you can identify forward-looking statements by terms such as “may,” “will,” “should,” “expect,” “plan,” “aim,” “anticipate,” “could,” “intend,” “target,” “project,” “contemplate,” “believe,” “estimate,” “predict,” “potential” or “continue” or the negative of these terms or other similar expressions. The forward-looking statements in this presentation are only predictions. The Company has based these forward-looking statements largely on its current expectations and projections about future events and financial trends that it believes may affect the Company’s business, financial condition and results of operations. These forward-looking statements speak only as of the date of this presentation and are subject to a number of risks, uncertainties and assumptions, including those listed under Risk Factors in the Company’s Form 10-K and other filings with the SEC, some of which cannot be predicted or quantified and some of which are beyond the Company’s control. The events and circumstances reflected in the Company’s forward-looking statements may not be achieved or occur, and actual results could differ materially from those projected in the forward-looking statements. Moreover, the Company operates in a dynamic industry and economy. New risk factors and uncertainties may emerge from time to time, and it is not possible for management to predict all risk factors and uncertainties that the Company may face. Except as required by applicable law, the Company does not plan to publicly update or revise any forward-looking statements contained herein, whether as a result of any new information, future events, changed circumstances or otherwise.

# Agenda

PRESENTATION	SPEAKER
<b>Welcome &amp; Introduction</b>	<b>Kate Rausch</b> Vice President, Investor Relations & Corporate Affairs
<b>Wave Evolution and Growth Drivers</b>	<b>Paul Bolno, MD, MBA</b> President and Chief Executive Officer
<b>GSK Perspectives: An Inflection Point with RNA Medicines</b>	<b>Tony Wood, PhD</b> Chief Scientific Officer, GSK
	<b>Carolyn Buser-Doepner, PhD</b> Vice President, Novel Human Genetics Research Unit, GSK
<b>RNAi: INHBE and Beyond</b>	<b>Chandra Vargeese, PhD</b> Chief Technology Officer
<b>AIMers: Editing to Upregulate</b>	<b>Chandra Vargeese, PhD</b> Chief Technology Officer
<b>Mapping the “Edit-verse”</b>	<b>Kenneth Longo, PhD</b> Vice President, Data Science
<b>Mining the “Edit-verse”</b>	<b>Ginnie Yang, PhD</b> Senior Vice President, Translational Medicine
<b>Closing Remarks</b>	<b>Paul Bolno, MD, MBA</b> President and Chief Executive Officer
<b>Q&amp;A</b>	<b>Wave Leadership Team</b>

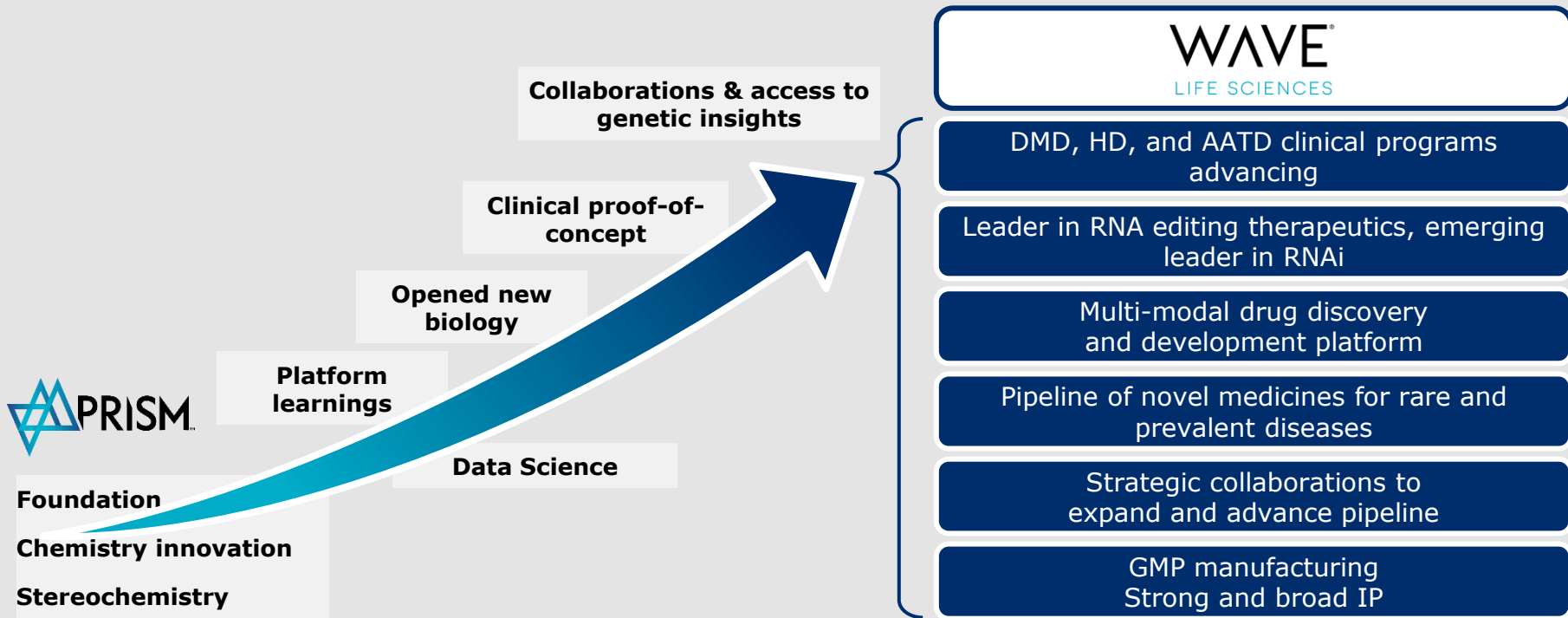


# Wave Evolution and Growth Drivers

Paul Bolno, MD, MBA  
*President and CEO*

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# Wave today is well positioned for significant and sustained growth



# Data from DMD and HD programs demonstrate clinical translation of Wave's platform chemistry

**DMD – WVE-N531**

High tissue concentrations and highest reported exon-skipping with three biweekly doses

**HD – WVE-003**

Most advanced allele-selective approach, with reductions in mHTT, preservation of wtHTT with single dose

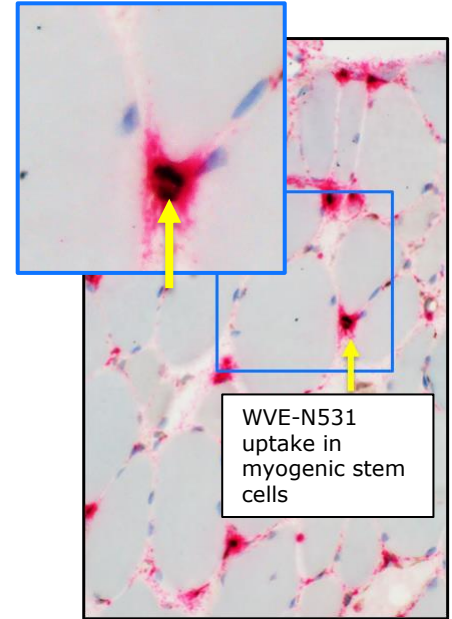
**AATD – WVE-006**

Clinical development initiated on the first RNA editing medicine

# WVE-N531 Part A data in DMD: High exon-skipping & muscle concentrations after three bi-weekly doses

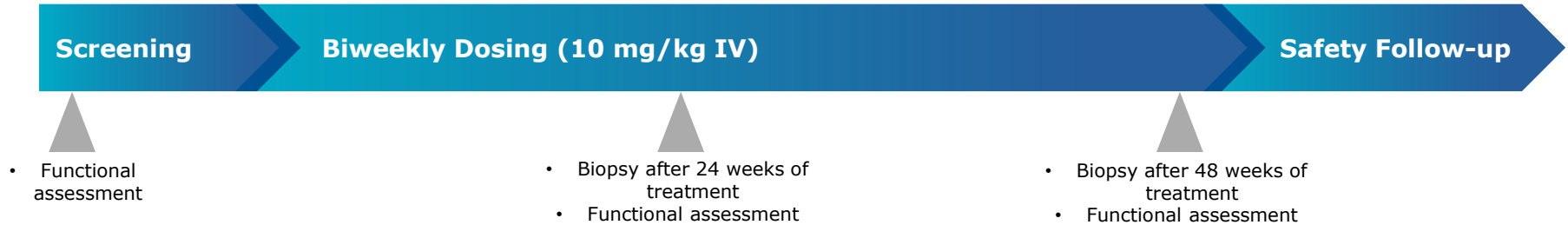
	suvidirsen	WVE-N531
Mean muscle concentration	0.7 $\mu\text{g/g}$	42 $\mu\text{g/g}$
Mean exon skipping	Not detectable	53%
Half-life in plasma	18 hours	25 days
Dose	22 weekly doses of 5 mg/kg	3 biweekly doses of 10 mg/kg

## WVE-N531 uptake in myocyte stem cells



Important for potential muscle regeneration

# FORWARD-53, a potentially registrational Phase 2 clinical trial of WVE-N531 in DMD (Exon 53)



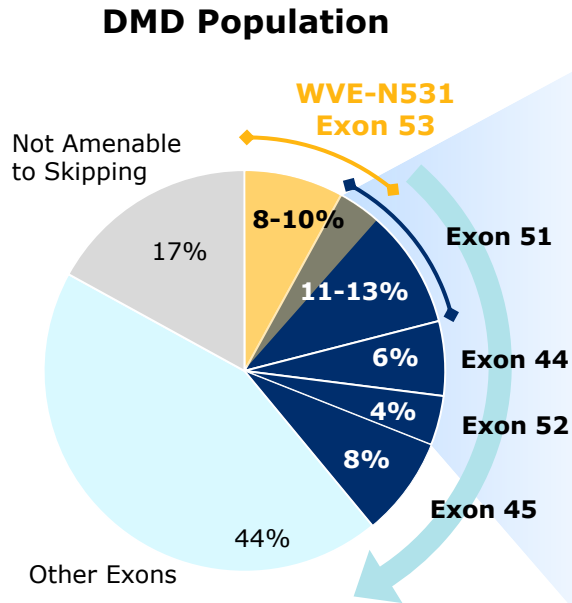
- **Design of FORWARD-53:** Phase 2, open-label, 10 mg/kg every other week, up to 10 patients
- **Endpoints:** Dystrophin (powered for >5% of normal), safety/tolerability, pharmacokinetics, functional assessments (incl. NSAA and others)
- **Biopsies:**
  - After 24 weeks of treatment
  - After 48 weeks of treatment



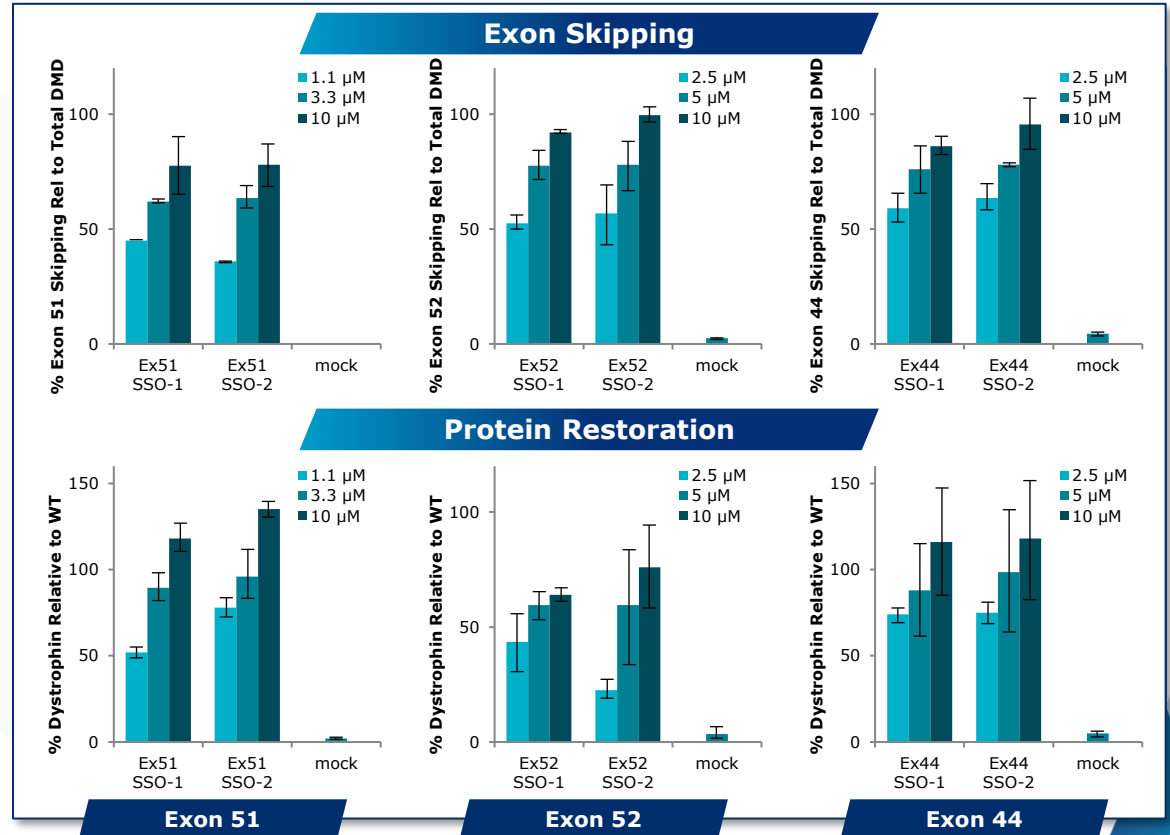
Data from FORWARD-53 expected in 2024



# Potential for Wave to address up to ~40% of DMD population

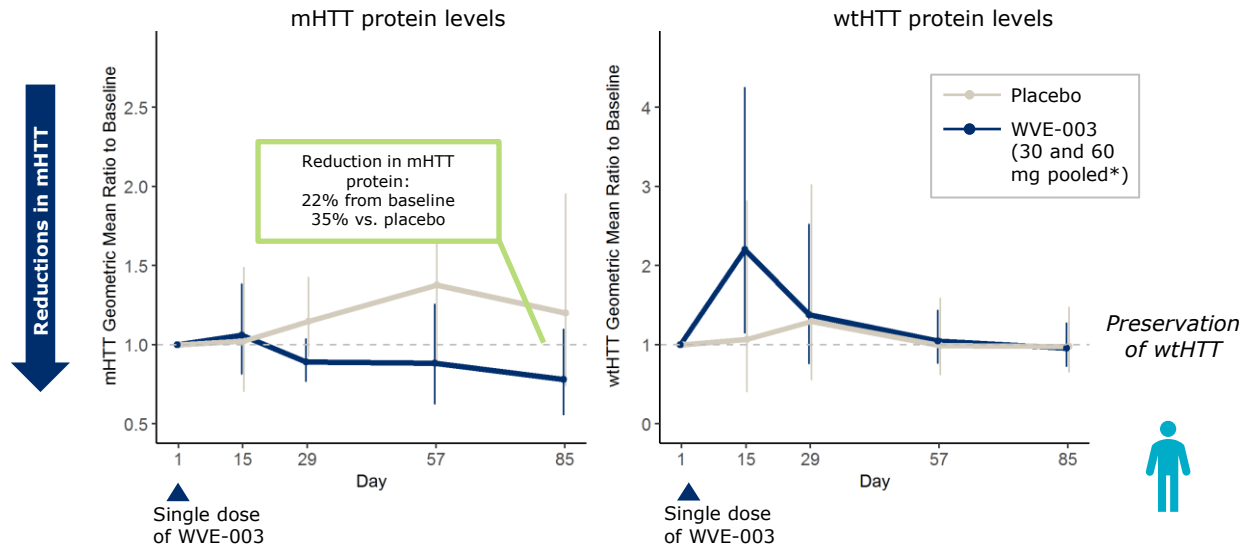


## Exon skipping and dystrophin restoration demonstrated *in vitro*



# WVE-003 in HD: Reductions in mHTT and preservation of wtHTT in single dose cohorts; multidose underway

**Reductions in mean CSF mHTT and preservation of wtHTT observed in pooled analysis of single dose cohorts in SELECT-HD clinical study**



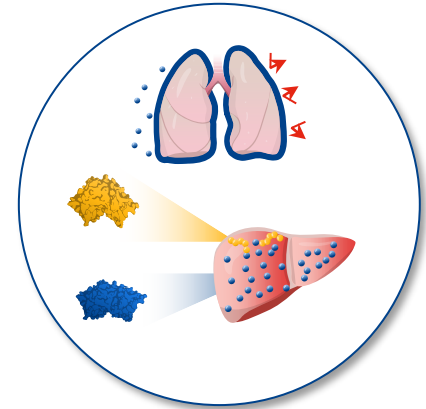
**Additional single-dose and available multi-dose data expected in 2H 2023  
Complete multi-dose data from first cohort with extended follow-up expected 2Q 2024**

# Preclinical data support WVE-006 as comprehensive approach to address AATD-related lung and liver disease

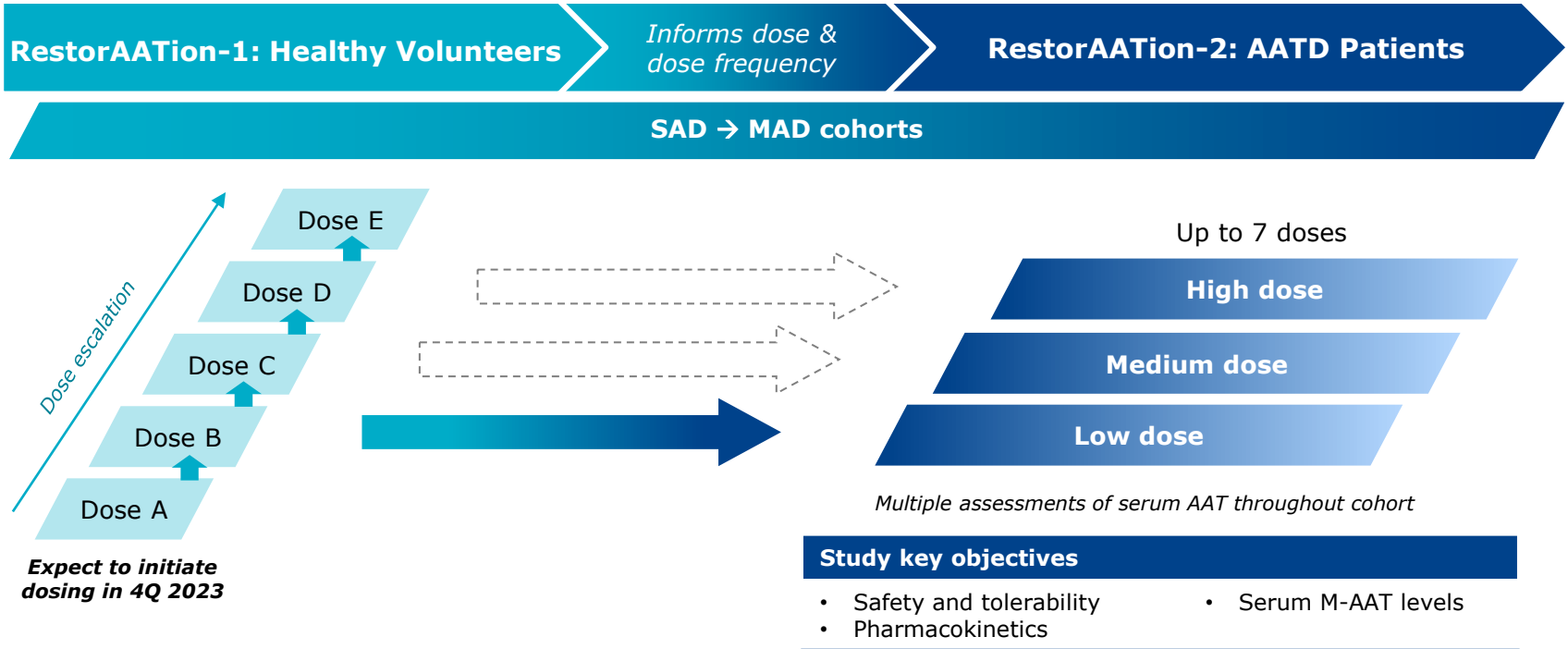


## Achieved key treatment goals with preclinical *in vitro* and *in vivo* datasets:

- ✓ **Significant increase in serum AAT of up to 30 uM in NSG-PiZ mice**
  - ~50% editing supports restoration to MZ phenotype
- ✓ **Restored wild-type M-AAT protein**
  - ~50% of AAT protein in serum is wild-type M-AAT
- ✓ **Editing is highly specific**
  - No bystander edits
- ✓ **Functionality of M-AAT protein**
  - >3-fold improvement in neutrophil elastase inhibition activity
- ✓ **Improvement in liver phenotype**
  - Decreased lobular inflammation and PAS-D globule size, prevents increase in hepatocyte turnover

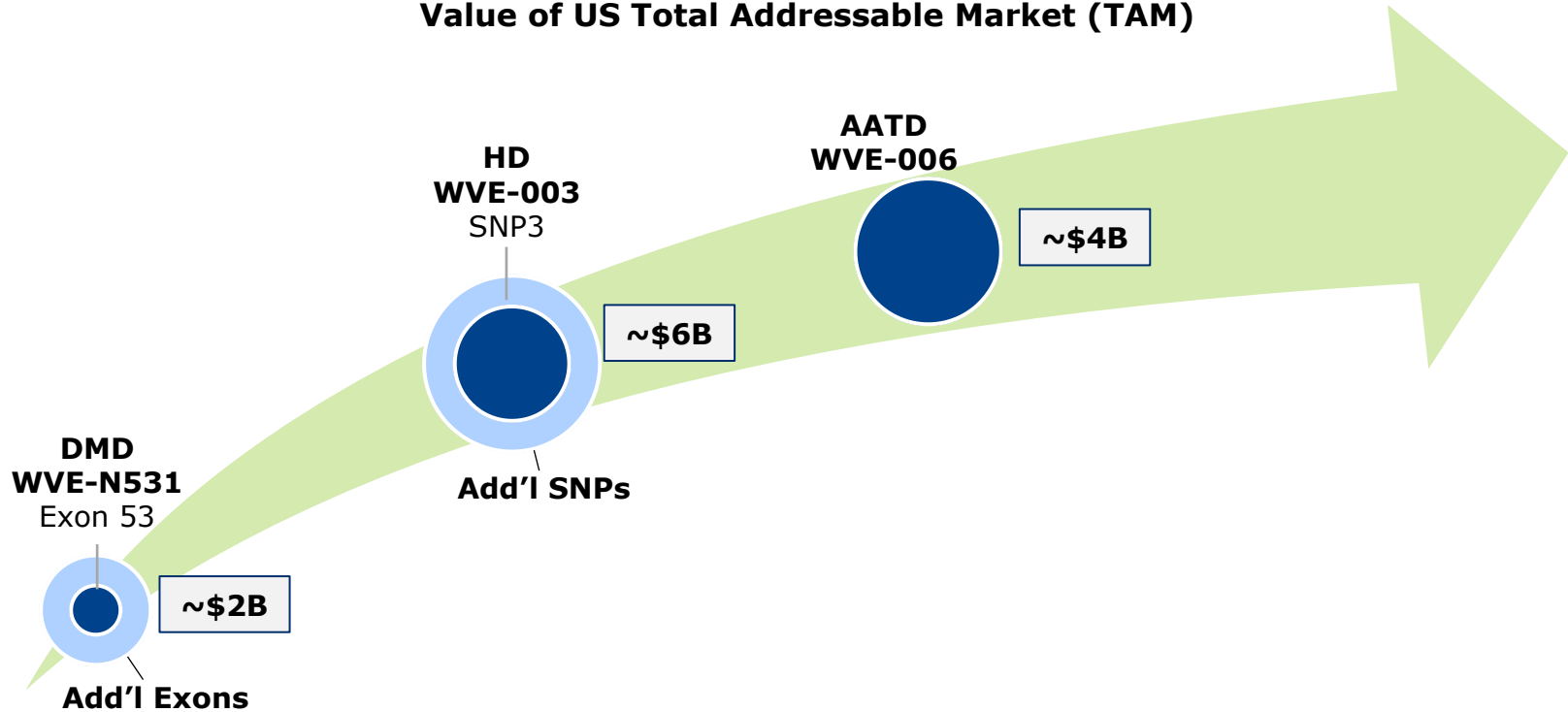


# Proof of mechanism data in patients with AATD expected in 2024

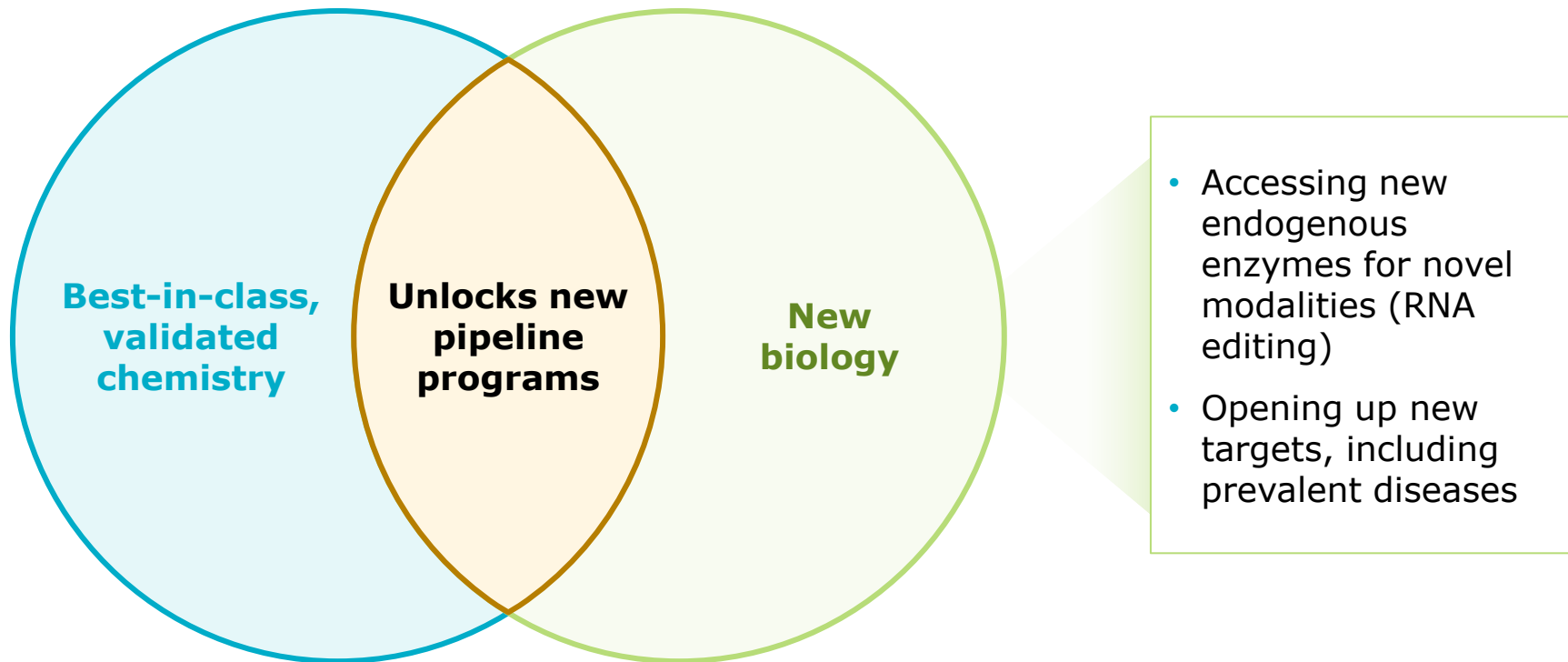


# Wave's current clinical programs represent significant market opportunity

## Value of US Total Addressable Market (TAM)



# Combining novel biology with validated, best-in-class chemistry to open opportunities for first-in-class medicines



# RNA editing enables correction of driver mutations as well as modulation of RNA and downstream proteins

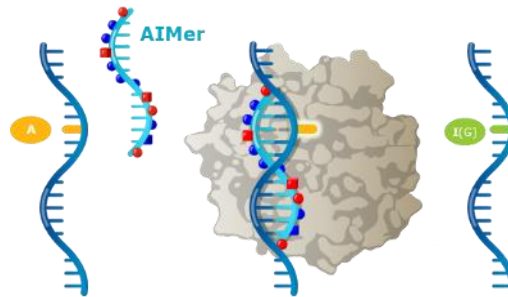
Correct G-to-A driver mutations with AIMers

Modulate protein expression with AIMers



Restore or correct protein function

**WVE-006**  
**(GalNAc AIMER)**  
AATD



**Upregulate expression to increase endogenous protein activity**

Applications to address rare and common diseases

- Mutation-agnostic approach to increase protein levels in diseases resulting from loss of protein function
- Increase endogenous production of therapeutic proteins

# Potential to address prevalent diseases with RNA editing upregulation

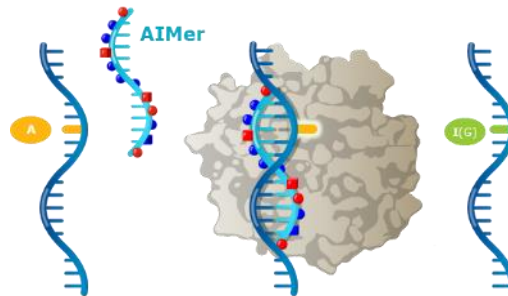
Correct G-to-A driver mutations with AIMers

Modulate protein expression with AIMers



Restore or correct protein function

**WVE-006**  
(GaNAC AIMER)  
AATD

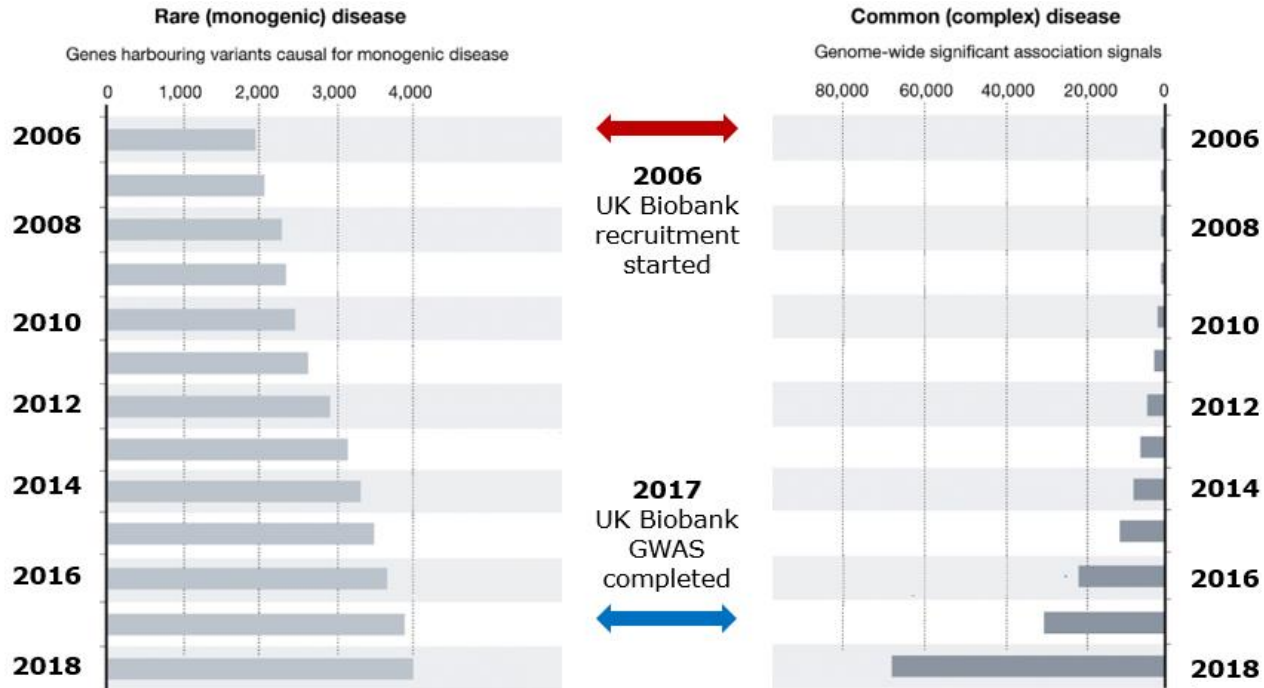


Upregulate expression to increase endogenous protein activity

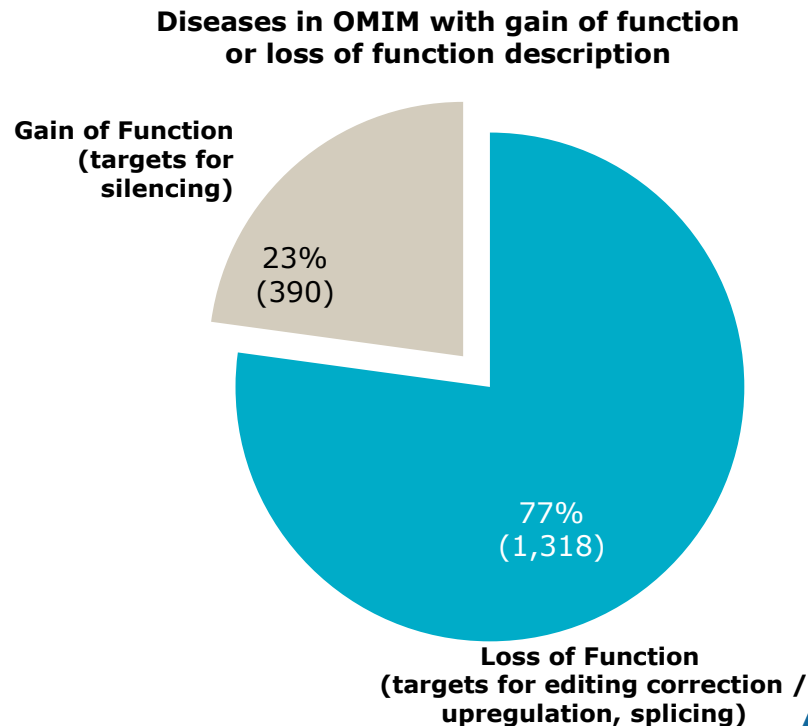
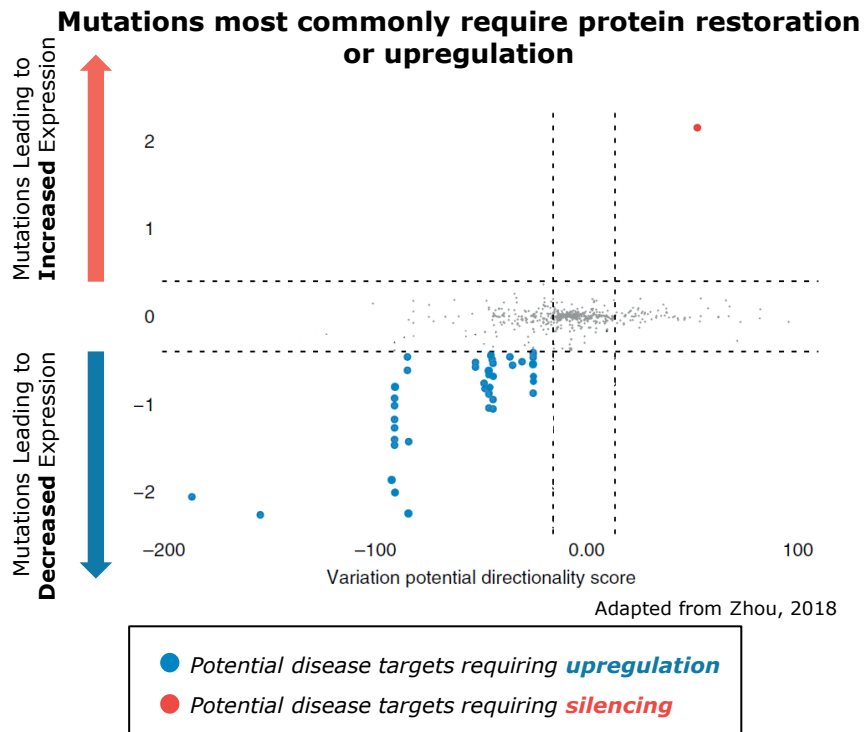
Potential to address prevalent diseases



# Increasing genetic insights for rare and common diseases is unlocking new target opportunities

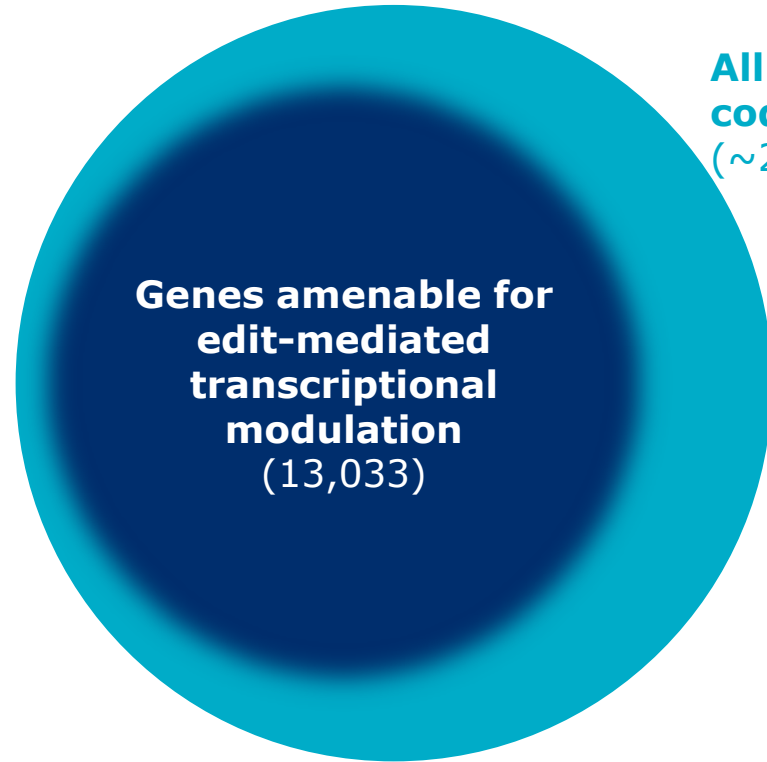


# Majority of disease associated mutations are predicted to decrease protein expression



# The AIMer-targetable 'Edit-Verse' is substantial











- **The Edit-verse is the editable gene-disease universe, including upregulation**
- **>13,000 genes** with a high-probability<sup>1</sup> of being amenable to transcriptional regulation with A-to-G editing
- Model development ongoing to expand access to **more protein-coding genes** and expand the Edit-verse



**All protein coding genes**  
(~22,000)

● **Expanding 'Edit-verse'**

# Robust RNA medicines pipeline with five new clinical candidates by year-end 2025

Program	Discovery	Preclinical	Clinical	Rights	Patient population (US & Europe)
<b>RNA EDITING</b>					
<b>WVE-006</b> SERPINA1 (AATD)				<b>GSK exclusive global license</b>	<b>200K</b>
Multiple undisclosed Correction				<b>100% global</b>	<b>&gt;20K (multiple)</b>
Multiple undisclosed Upregulation				<b>100% global</b>	<b>&gt;3M (multiple)</b>
<b>SPLICING</b>					
<b>WVE-N531</b> Exon 53 (DMD)				<b>100% global</b>	<b>2.3K</b>
Other exons (DMD)				<b>100% global</b>	<b>Up to 18K</b>
<b>SILENCING: ANTISENSE</b>					
<b>WVE-003</b> mHTT (HD)				<b>Takeda 50:50 Option</b>	<b>25K Manifest (SNP3) 60K Pre-Manifest (SNP3)</b>
<b>SILENCING: RNAi</b>					
<b>INHBE*</b> (Metabolic disorders)				<b>100% global</b>	<b>47M</b>



# GSK Perspectives: An Inflection Point with RNA Medicines



Tony Wood, PhD  
*Chief Scientific Officer, GSK*

Carolyn Buser-Doepner, PhD  
*Vice President, Novel Human Genetics  
Research Unit, GSK*

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# **GSK Perspectives: An Inflection Point with RNA Medicines**

Tony Wood, PhD  
*Chief Scientific Officer, GSK*

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# **GSK Perspectives: An Inflection Point with RNA Medicines**

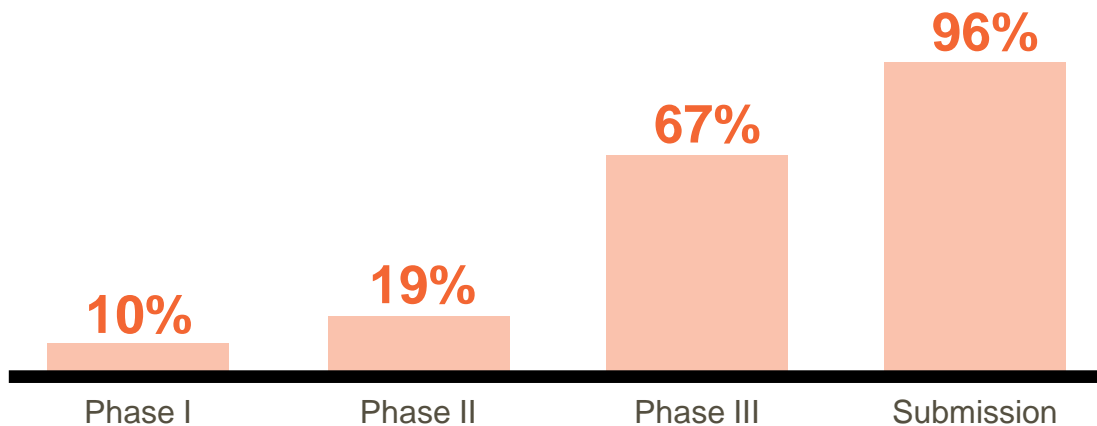
Carolyn Buser-Doepner, PhD  
*Vice President, Novel Human Genetics  
Research Unit, GSK*

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# Biggest challenge of drug development:

90% of potential medicines that enter the clinic FAIL!

Project success rates to approval  
2015-2019



## Why?

Traditional target selection has been flawed

- Animal models not predictive
- Correlation vs. causation



# Genetic evidence supports drug development

Drugs with human genetic evidence are >2x more likely to succeed

<u>V2G method</u>	<u>pValue</u>	<u>Odds ratio</u>
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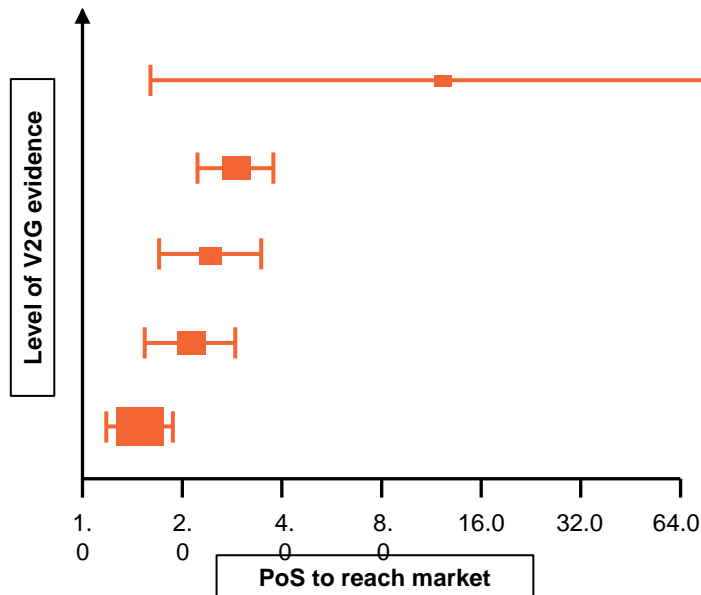
pLoF	4.8e-03	9.95
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OMIM	2.0e-30	3.14
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missense	2.1e-11	2.77
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eQTL/pQTL	4.4e-11	2.31
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nearest gene	5.1e-08	1.55
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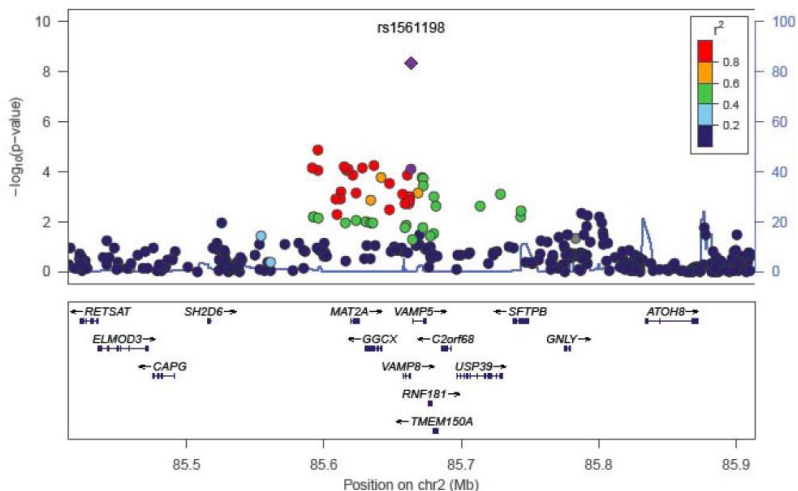


Drugs with human genetic evidence are >2x more likely to succeed

PoS proportional to strength of evidence for causality

# Genetic association is just the first step

More challenging to identify the causal gene and its function (V2G2F)



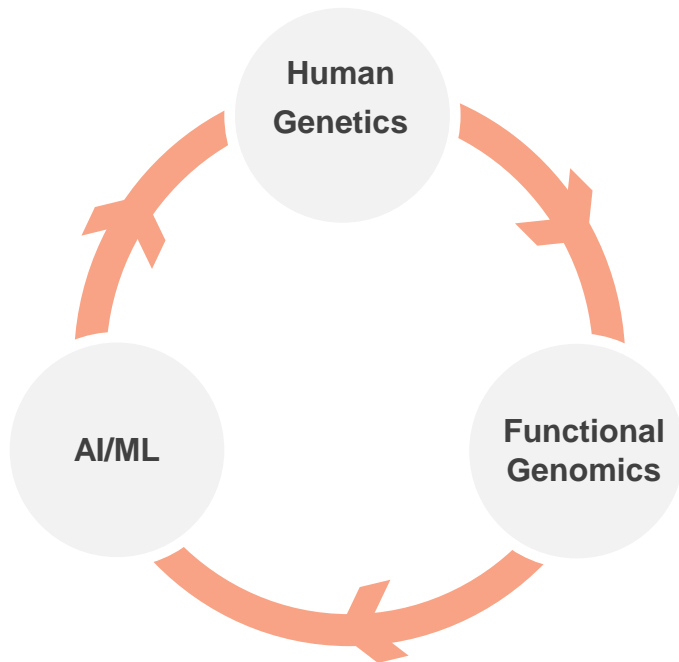
**Need for experimental "wet" work to validate *in silico* associations**

- Fine mapping (sequencing)
- Expression (eQTL / pQTL)
- Other molecular features (chromatin structure & interactions)
- Gene perturbation (CRISPR, TALE)

Multiple candidate genes in region  
>95% of GWAS variants outside of exons

# GSK has built competitive advantage for target identification

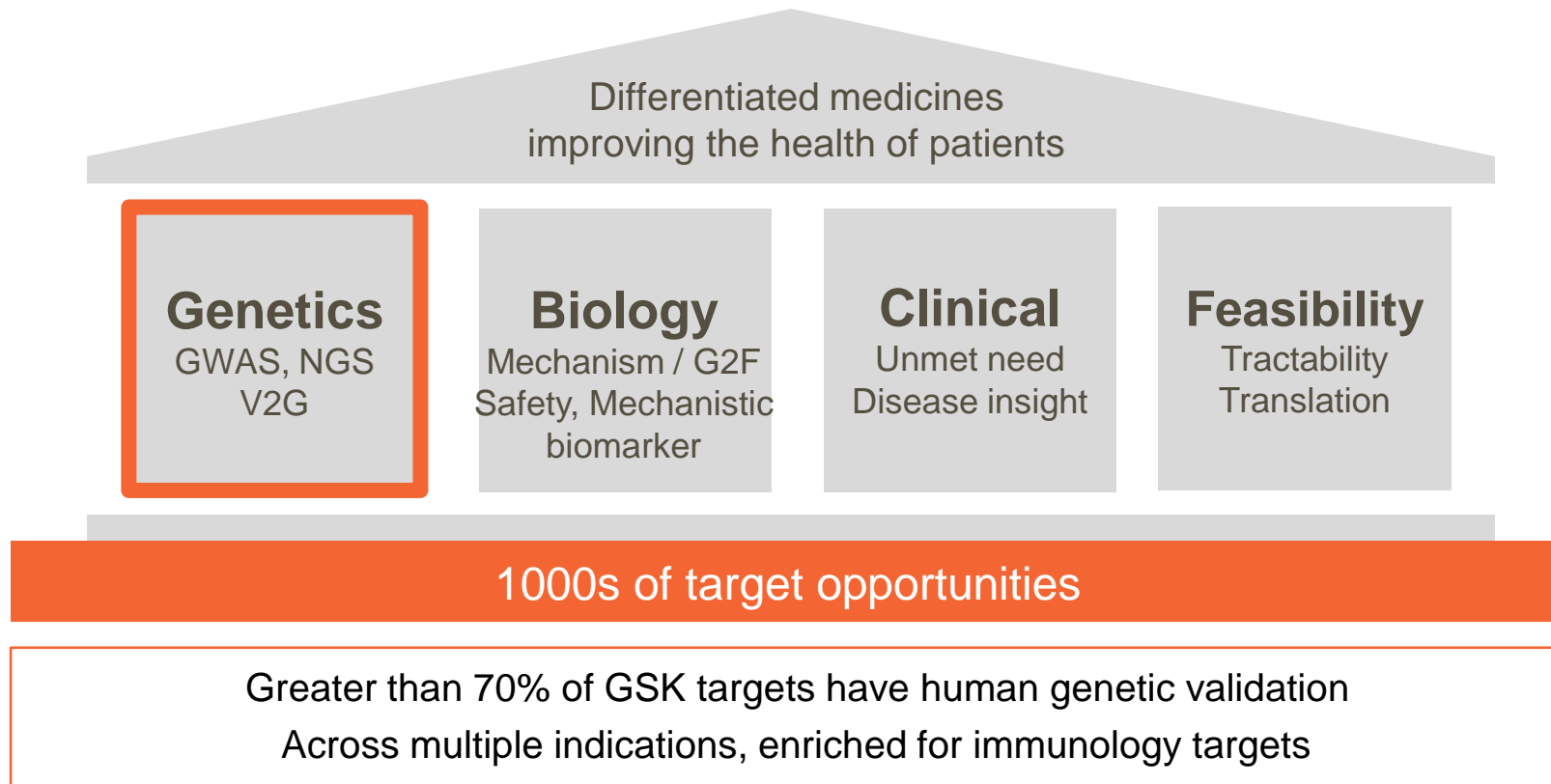
Mapping of genetic signals points towards modality to regulate RNA



## Rationale for RNA oligo therapy

- >90% of genetic signals from GWAS map to non-coding elements
- Modulation of proteome
- Modulation of regulome

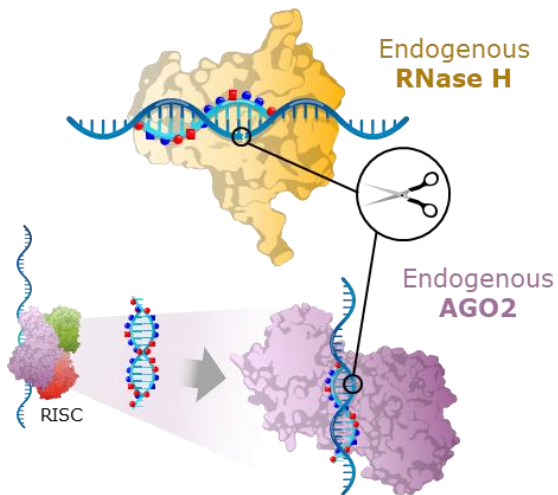
# Pillars underlying target identification and prioritization



# Wave's best-in-class multi-modal platform enables silencing, splicing and editing modalities

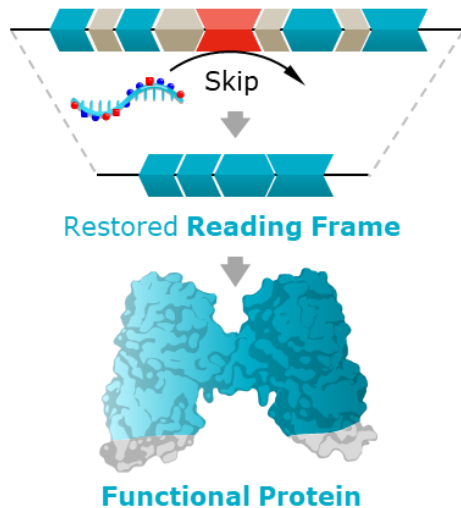
## Silencing

- Degrade RNA transcripts to **turn off** protein production



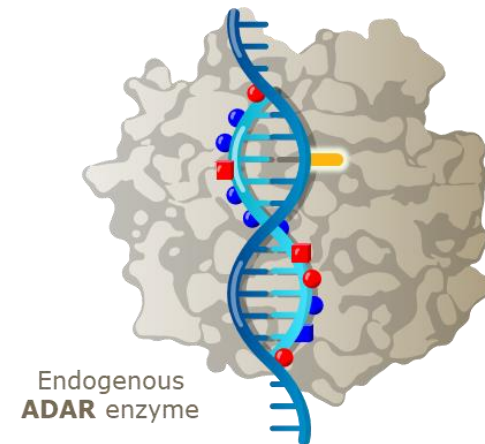
## Splicing

- Restore RNA transcripts and **turn on** protein production



## RNA Base Editing

- Edit RNA bases to **restore** or **modulate** protein production





# RNAi: INHBE and Beyond

Chandra Vargeese, PhD  
*Chief Technology Officer*

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# Multiple recent publications on Wave leadership in RNA therapeutics



## Platform

### CNS silencing

Variant-selective stereorec oligonucleotide protect against pC9orf7-repeat ex

Published online 2 February 2023

<https://doi.org/10.1038/s41587-022-01047-7>

**NAR Breakthrough Article**  
**Impact of guanidine-containing backbone on stereorec antisense oligonucleotides in**

Pachamuthu Kandamany<sup>1</sup>, Xianying Liu<sup>1</sup>, Vincent Adida, Sandesh Amy Andreucci, David Boulay, Keith Bonome, Michael Byrne, Mounir Chahine, Chandra Vargese<sup>1,2</sup>, Erin Purcell Estabrook, Jullip Dilip Sheike, Himani Shah, Anthony Lamattina, Gianni Pan, Brett Schwanz, Frank Favalaro, Mugisha Bedekar, Arindom Chatterjee, Jigar Desai<sup>1,2</sup>, Tomomi Kawamoto, Gentang Lu, Jake Mettenville, Milinda Sarawara, Priyanka Shiva Prakashia, Hailin Yang, Yuan Yin, Hai Ya, Paloma H. Giangrande, Michael Byrne, Pachamuthu Kandamany and Chandra Vargese<sup>1,2</sup>

Waves Life Sciences, Cambridge, MA 02138, USA

Received May 30, 2022; Revised December 17, 2022; Accepted 1

**ABSTRACT**  
 While recent advances in antisense oligonucleotide (ASO) technology have enabled the development of ASOs for the treatment of a wide range of diseases, the development of ASOs for the treatment of neurodegenerative diseases remains a challenge. Here, we report the synthesis and impact of stereorec oligonucleotides containing a guanidine backbone (GASOs) on the efficacy of ASOs in a mouse model of Huntington's disease. We show that GASOs exhibit improved efficacy compared to ASOs with a sugar-phosphate backbone. We also show that GASOs exhibit improved efficacy compared to ASOs with a sugar-phosphate backbone in a mouse model of Huntington's disease.

**INTRODUCTION**  
 The development of RNA therapeutics (RTs) has been a major focus of research in the pharmaceutical industry. RTs include antisense oligonucleotides (ASOs), small interfering RNAs (siRNAs), and messenger RNA (mRNA) vaccines. ASOs are short, single-stranded nucleic acid molecules that bind to a target RNA sequence and inhibit its function. ASOs have been used to treat a variety of diseases, including cancer, genetic disorders, and neurodegenerative diseases. However, the development of ASOs for the treatment of neurodegenerative diseases remains a challenge. One of the major challenges is the low efficacy of ASOs in the brain. This is due to the low permeability of ASOs across the blood-brain barrier (BBB). To overcome this challenge, researchers have developed various strategies to improve the efficacy of ASOs in the brain. One such strategy is the use of stereorec oligonucleotides (ASOs) with a guanidine backbone (GASOs). GASOs are ASOs that contain a guanidine group in the backbone. This group is thought to improve the permeability of ASOs across the BBB. We have previously reported the synthesis and impact of GASOs on the efficacy of ASOs in a mouse model of Huntington's disease. Here, we report the synthesis and impact of stereorec oligonucleotides containing a guanidine backbone (GASOs) on the efficacy of ASOs in a mouse model of Huntington's disease. We show that GASOs exhibit improved efficacy compared to ASOs with a sugar-phosphate backbone. We also show that GASOs exhibit improved efficacy compared to ASOs with a sugar-phosphate backbone in a mouse model of Huntington's disease.

## siRNA

Published online 18 April 2023

### Impact of stereorec chimeric backbone chemistries on the potency and durability of gene silencing by RNA interference

Wei Liu<sup>1</sup>, Naoki Iwamoto<sup>1</sup>, Subramanian Maraganan, Xue Lu, Snehaa Tripathi, Erin Purcell Estabrook, Jullip Dilip Sheike, Himani Shah, Anthony Lamattina, Gianni Pan, Brett Schwanz, Frank Favalaro, Mugisha Bedekar, Arindom Chatterjee, Jigar Desai<sup>1,2</sup>, Tomomi Kawamoto, Gentang Lu, Jake Mettenville, Milinda Sarawara, Priyanka Shiva Prakashia, Hailin Yang, Yuan Yin, Hai Ya, Paloma H. Giangrande, Michael Byrne, Pachamuthu Kandamany and Chandra Vargese<sup>1,2</sup>

Waves Life Sciences, Cambridge, MA 02138, USA

Received December 10, 2022; Revised March 30, 2023; Editorial Decision March 27, 2023; Accepted March 27, 2023

**ABSTRACT**  
 We report the systematic investigation of stereorec phosphorothioate (PS) and phosphorothioate (PT) linkages on siRNA-mediated silencing. The incorporation of appropriately positioned PS and PT linkages to the 3' end of the siRNA backbone improved silencing efficacy and durability of response, especially in a mouse model of Huntington's disease. We also show that the incorporation of PS and PT linkages to the 3' end of the siRNA backbone improved silencing efficacy and durability of response, especially in a mouse model of Huntington's disease. We also show that the incorporation of PS and PT linkages to the 3' end of the siRNA backbone improved silencing efficacy and durability of response, especially in a mouse model of Huntington's disease.

**INTRODUCTION**  
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## Splicing

Published online 21 January 2023

### Control of backbone chemistry and chirality boost oligonucleotide splice switching activity

Pachamuthu Kandamany<sup>1</sup>, Graham McClorey<sup>1,2</sup>, Mamoru Shimizu<sup>1</sup>, Nayantara Kothari<sup>1</sup>, Ronshah Aiam<sup>1</sup>, Naoki Iwamoto<sup>1</sup>, Jayakantham Kumarasamy<sup>1</sup>, Gopal R. Boonmoung<sup>1</sup>, Adam Basglin<sup>1</sup>, Onanong Chivatavakorn<sup>1</sup>, David C. D. Butler<sup>1</sup>, Michael Byrne<sup>1</sup>, Katarzyna Chwielewska<sup>1</sup>, Kay E. Davies<sup>1</sup>, Jigar Desai<sup>1,2</sup>, Jullip Dilip Sheike<sup>1</sup>, Ann F. Durbin<sup>1</sup>, Ruth Elkington<sup>1</sup>, Ben Edwards<sup>1</sup>, Jack Godfrey<sup>1</sup>, Andrew Hoes<sup>1</sup>, Fangfang Liu<sup>1</sup>, Kenneth Longo<sup>1</sup>, Gentang Lu<sup>1</sup>, Subramanian Maraganan<sup>1</sup>, Jacopo Oleni<sup>1</sup>, R. Hutton Paik<sup>1</sup>, Erin Purcell Estabrook<sup>1</sup>, Chikida Shivalla<sup>1</sup>, Wasee Taschian<sup>1</sup>, Tomomi Kawamoto<sup>1</sup>, Carlo Rinaldi<sup>1,2,3</sup>, Joana Rajao-Saravali<sup>1</sup>, Snehaa Tripathi<sup>1</sup>, Hailin Yang<sup>1</sup>, Yuan Yin<sup>1</sup>, Xianyi Zhao<sup>1</sup>, Cong Zhou<sup>1,2</sup>, Jason Zhang<sup>1</sup>, Luciano Apponi<sup>1</sup>, Matthew J. A. Wood<sup>1,2</sup> and Chandra Vargese<sup>1,2</sup>

Waves Life Sciences, Cambridge, MA, USA; <sup>2</sup>Department of Paediatrics, University of Oxford, South Parks Road, Oxford OX3 9DU, UK; <sup>3</sup>NIHR Oxford Neuroimaging Centre, University of Oxford, Oxford OX3 9DU, UK and <sup>4</sup>Department of Physics, Anatomy and Genetics, University of Oxford, South Parks Road, Oxford OX1 3PS, UK

Received September 01, 2021; Revised December 18, 2022; Editorial Decision January 10, 2023; Accepted January 07, 2023

**ABSTRACT**  
 Although recent regulatory approval of splice-switching oligonucleotides (SSOs) for the treatment of neurodegenerative disease such as Duchenne muscular dystrophy has been an advance for the application of neurochemicals, it has also highlighted the need for improved SSO chemistries to reach tissues such as the skeletal muscle and heart.

**INTRODUCTION**  
 RNA-splicing oligonucleotides, a type of applied nucleic acid, are designed to bind through complementary base pairing to pre-mRNA, causing the skipping, inclusion, or retention of exons in the splicing machinery and hence producing a specific protein isoform. RNA-splicing oligonucleotides (SSOs) have been used to treat a variety of diseases, including cancer, genetic disorders, and neurodegenerative diseases. However, the development of SSOs for the treatment of neurodegenerative diseases remains a challenge. One of the major challenges is the low efficacy of SSOs in the brain. This is due to the low permeability of SSOs across the blood-brain barrier (BBB). To overcome this challenge, researchers have developed various strategies to improve the efficacy of SSOs in the brain. One such strategy is the use of stereorec oligonucleotides (SSOs) with a guanidine backbone (GASOs). GASOs are SSOs that contain a guanidine group in the backbone. This group is thought to improve the permeability of SSOs across the BBB. We have previously reported the synthesis and impact of GASOs on the efficacy of SSOs in a mouse model of Huntington's disease. Here, we report the synthesis and impact of stereorec oligonucleotides containing a guanidine backbone (GASOs) on the efficacy of SSOs in a mouse model of Huntington's disease. We show that GASOs exhibit improved efficacy compared to SSOs with a sugar-phosphate backbone. We also show that GASOs exhibit improved efficacy compared to SSOs with a sugar-phosphate backbone in a mouse model of Huntington's disease.

**CONCLUSION**  
 We have shown that the incorporation of PS and PT linkages to the 3' end of the siRNA backbone improved silencing efficacy and durability of response, especially in a mouse model of Huntington's disease. We also show that the incorporation of PS and PT linkages to the 3' end of the siRNA backbone improved silencing efficacy and durability of response, especially in a mouse model of Huntington's disease.

## Editing

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### Endogenous ADAR-mediated RNA editing in non-human primates using stereorec chemically modified oligonucleotides

Prashant Manian<sup>1</sup>, Chikida Shivalla<sup>1</sup>, Gentang Lu<sup>1</sup>, Mamoru Shimizu<sup>1</sup>, David Boulay<sup>1</sup>, Karley Bonner<sup>1</sup>, Michael Byrne<sup>1</sup>, Adam Basglin<sup>1</sup>, Arindom Chatterjee<sup>1</sup>, David Chew<sup>1</sup>, Jigar Desai<sup>1,2</sup>, Frank Favalaro<sup>1</sup>, Jack Godfrey<sup>1</sup>, Andrew Hoes<sup>1</sup>, Naoki Iwamoto<sup>1</sup>, Tomomi Kawamoto<sup>1</sup>, Jayakantham Kumarasamy<sup>1</sup>, Anthony Lamattina<sup>1</sup>, Amber Lindsey<sup>1</sup>, Fangfang Liu<sup>1</sup>, Richard Looby<sup>1</sup>, Subramanian Maraganan<sup>1</sup>, Jake Mettenville<sup>1</sup>, Rosalind Murphy<sup>1</sup>, Jeff Roush<sup>1</sup>, Tim P. Bily Buttner<sup>1,2</sup>, Stephany Standley<sup>1</sup>, Snehaa Tripathi<sup>1</sup>, Hailin Yang<sup>1</sup>, Yuan Yin<sup>1</sup>, Hai Ya, Cong Zhou<sup>1,2</sup>, Luciano H. Apponi<sup>1</sup>, Pachamuthu Kandamany<sup>1</sup> and Chandra Vargese<sup>1,2</sup>

Waves Life Sciences, Cambridge, MA, USA; <sup>2</sup>Department of Paediatrics, University of Oxford, South Parks Road, Oxford OX3 9DU, UK and <sup>3</sup>NIHR Oxford Neuroimaging Centre, University of Oxford, Oxford OX3 9DU, UK

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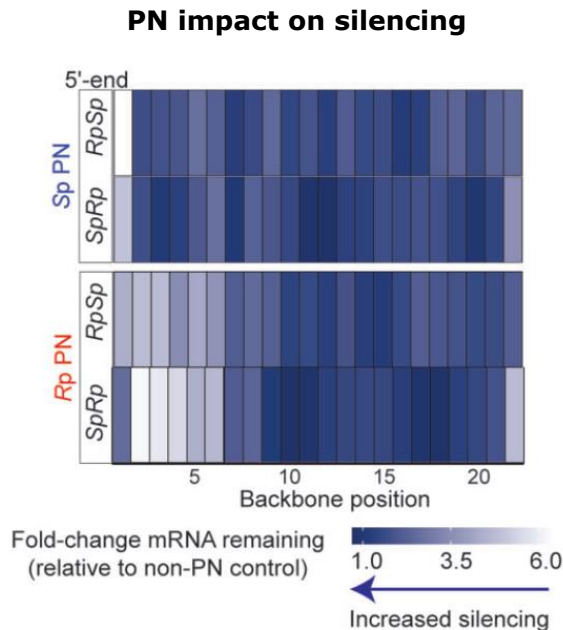
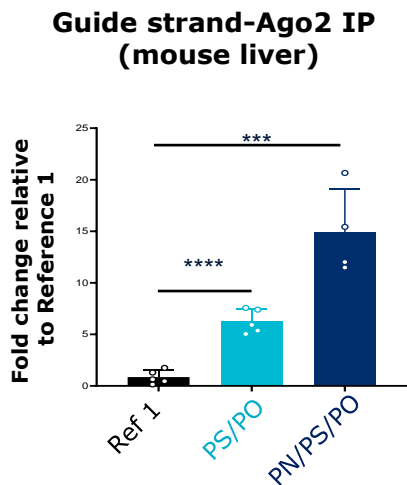
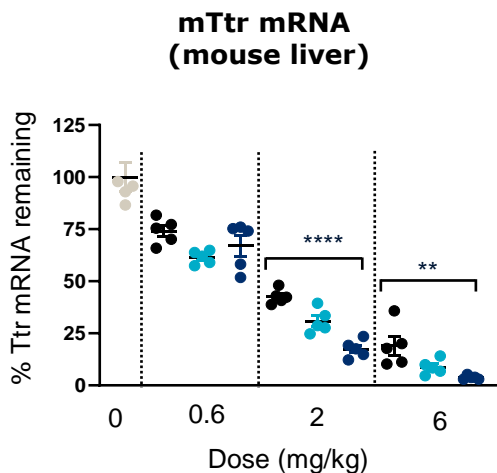
**ABSTRACT**  
 Although recent regulatory approval of splice-switching oligonucleotides (SSOs) for the treatment of neurodegenerative disease such as Duchenne muscular dystrophy has been an advance for the application of neurochemicals, it has also highlighted the need for improved SSO chemistries to reach tissues such as the skeletal muscle and heart.

**INTRODUCTION**  
 RNA-splicing oligonucleotides, a type of applied nucleic acid, are designed to bind through complementary base pairing to pre-mRNA, causing the skipping, inclusion, or retention of exons in the splicing machinery and hence producing a specific protein isoform. RNA-splicing oligonucleotides (SSOs) have been used to treat a variety of diseases, including cancer, genetic disorders, and neurodegenerative diseases. However, the development of SSOs for the treatment of neurodegenerative diseases remains a challenge. One of the major challenges is the low efficacy of SSOs in the brain. This is due to the low permeability of SSOs across the blood-brain barrier (BBB). To overcome this challenge, researchers have developed various strategies to improve the efficacy of SSOs in the brain. One such strategy is the use of stereorec oligonucleotides (SSOs) with a guanidine backbone (GASOs). GASOs are SSOs that contain a guanidine group in the backbone. This group is thought to improve the permeability of SSOs across the BBB. We have previously reported the synthesis and impact of GASOs on the efficacy of SSOs in a mouse model of Huntington's disease. Here, we report the synthesis and impact of stereorec oligonucleotides containing a guanidine backbone (GASOs) on the efficacy of SSOs in a mouse model of Huntington's disease. We show that GASOs exhibit improved efficacy compared to SSOs with a sugar-phosphate backbone. We also show that GASOs exhibit improved efficacy compared to SSOs with a sugar-phosphate backbone in a mouse model of Huntington's disease.

**CONCLUSION**  
 We have shown that the incorporation of PS and PT linkages to the 3' end of the siRNA backbone improved silencing efficacy and durability of response, especially in a mouse model of Huntington's disease. We also show that the incorporation of PS and PT linkages to the 3' end of the siRNA backbone improved silencing efficacy and durability of response, especially in a mouse model of Huntington's disease.




# Incorporation of PN modification improves *Ttr* GalNAc-siRNA in mice



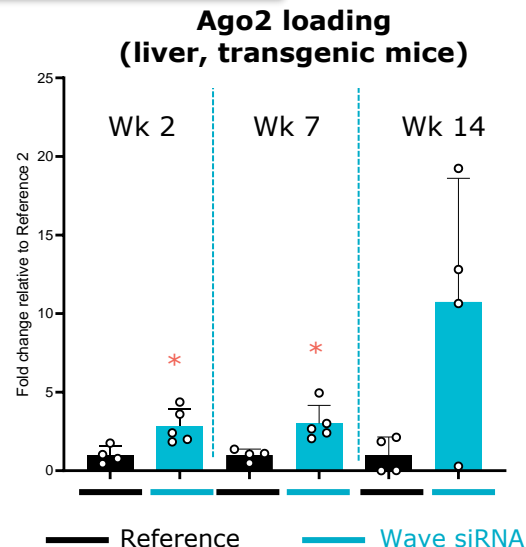
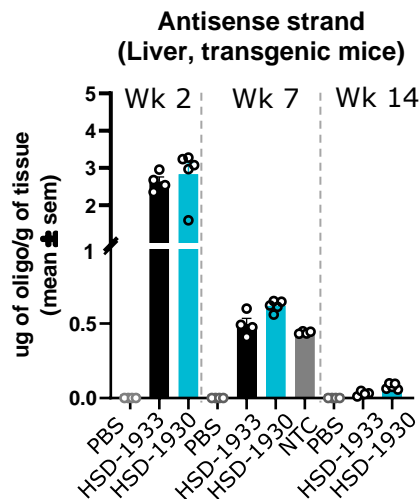
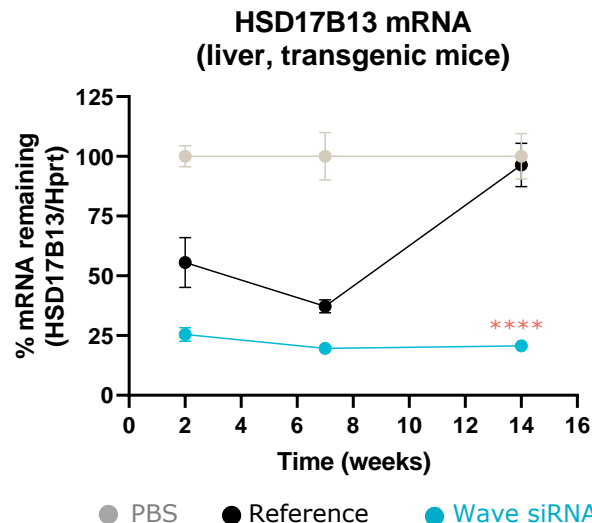
- PS/PO/PN backbone
- PS/PO backbone
- Reference compound
- PBS



# Potential for best-in-class RNAi enabled by Wave's PRISM platform

 Nucleic Acids Research  
Impact of stereopure chimeric backbone chemistries on the potency and durability of gene silencing by RNA interference

- Unprecedented Ago2 loading increases potency and durability of silencing following administration of single subcutaneous dose



RNAi is one of multiple Wave modalities being advanced in strategic research collaboration with GSK

# Driven by clinical genetics, Wave's first RNAi program addresses high unmet need in metabolic disorders, including obesity

## **INHBE program is Wave's first wholly owned program emerging from GSK collaboration**

- Leverages novel genetic insights accessed through GSK collaboration
- INHBE loss-of-function heterozygous carriers exhibit healthy metabolic profile<sup>1,2,3</sup>:
  - ✓ Reduced waist-to-hip circumference
  - ✓ Reduced odds ratio of Type 2 diabetes by 28%, and coronary artery disease
  - ✓ Reduced serum triglycerides
  - ✓ Elevated HDL-c
  - ✓ Reduced HbA1c
  - ✓ Lowered ApoB
- INHBE expressed primarily in liver and gene product (subunit of activin E) acts on its receptor in adipose tissue<sup>4</sup>
- GalNAc-siRNA for targeted delivery to hepatocytes

**≥50% reduction of INHBE with siRNA expected to restore a healthy metabolic profile**

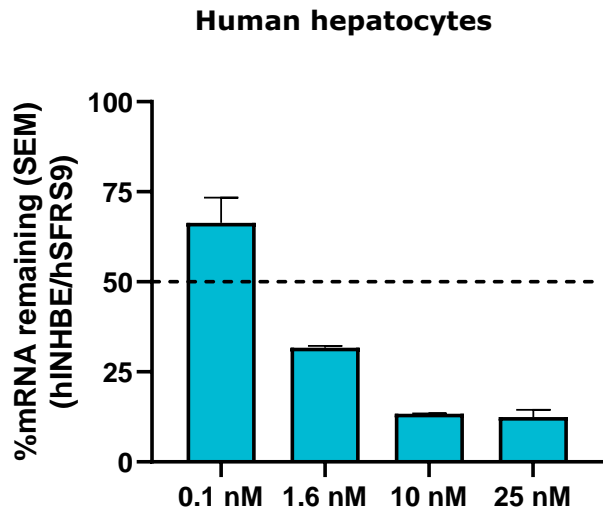
# INHBE GalNAc-siRNA represents an evolution in treatment for metabolic diseases, including obesity

- Metabolic syndrome\* is associated with type 2 diabetes, cardiovascular disease, hypertension, stroke, cancer, and increased mortality<sup>1,2</sup>
- Estimate ~47M people in US and Europe with metabolic disorders, including obesity
- Therapeutic options beyond GLP1s are needed
  - GLP1 receptor agonists lead to weight loss at the expense of muscle<sup>4</sup>
  - GLP1 receptor agonists suppress general reward system<sup>7</sup>
  - GLP1 receptor agonists associated with poor tolerability profile<sup>5</sup> with 68% drop-off after 1 year<sup>6</sup>
- Preferred approach would improve metabolism and increase fat loss while maintaining muscle mass
- Restoration of metabolic health via INHBE silencing can simultaneously address obesity and other drivers of metabolic syndrome

\*Patients diagnosed with metabolic syndrome based on having 3 of the following: abdominal obesity, high bp, high blood glucose, high TG, or low HDL

1. Liang, et al. 2023 Postgraduate Medical Journal 99(1175):985; 2. Lakka, et al. 2002 JAMA 288(21):2709; 3. Ryan and Yockey 2017 Curr Obes Rep 6(2):187; 4. Sargeant, et al. 2019 Endocrinol Metab (Seoul) 34(3):247-262; 5. Liu, et al. 2022 Front. Endocrinol. 13:1043789; 6. Prime Therapeutics Claims Analysis, July 2023; 7. Müller, et al. 2019 Molecular Metabolism 30: 72-130.

# INHBE knockdown of 90% demonstrated in human hepatocytes with GalNAc-siRNA

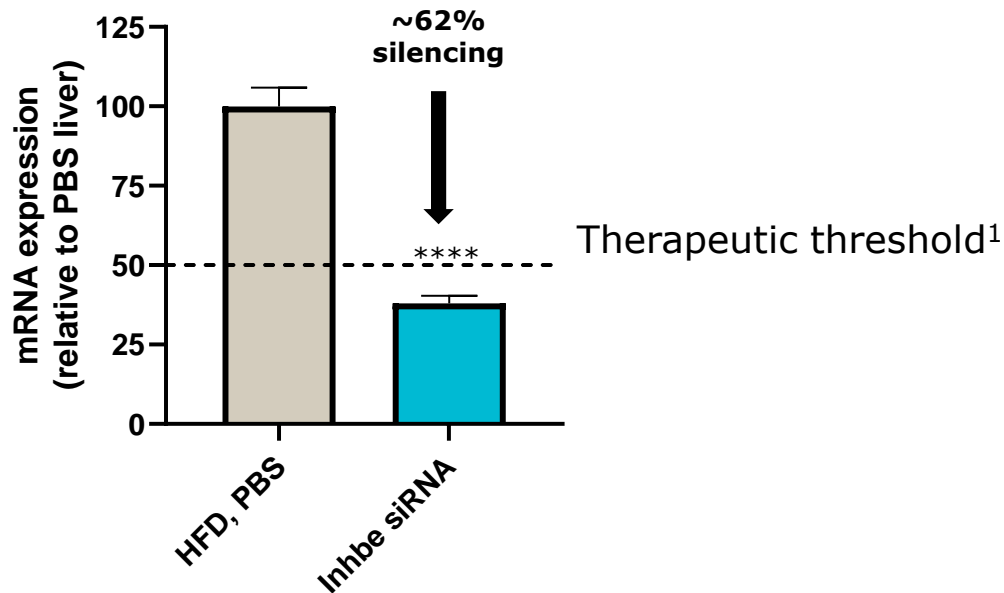


- This cross-reactive sequence demonstrates ~90% maximal knock-down in human hepatocytes and ~65% in mouse hepatocytes
- Additional human selective sequences are in development

# INHBE silencing achieved *in vivo* with GalNAc-siRNA exceeds therapeutic threshold

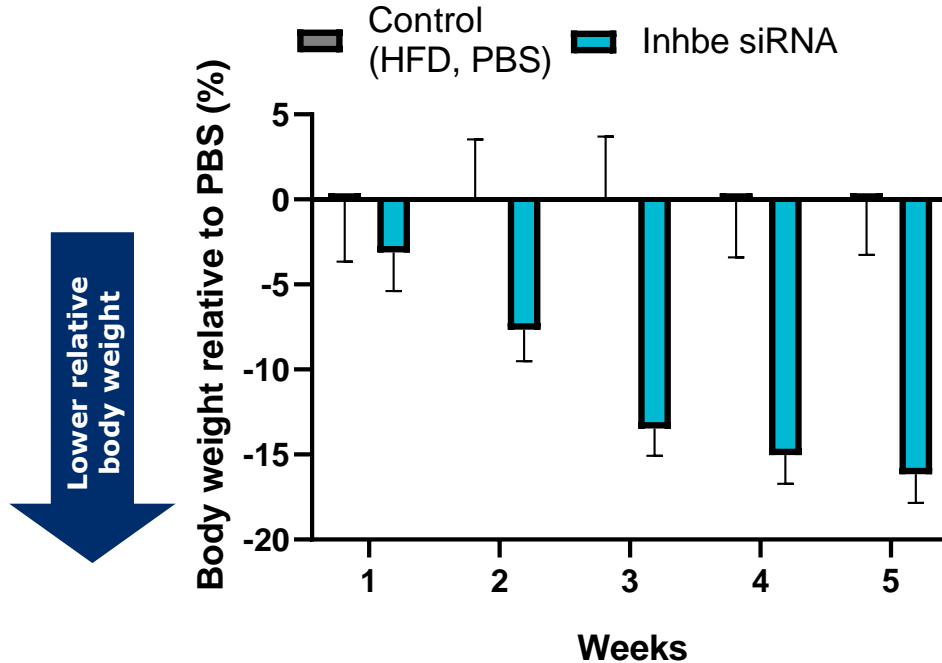


**INHBE knockdown demonstrated in mice at 5 weeks**



# INHBE knockdown led to 16% lower body weight

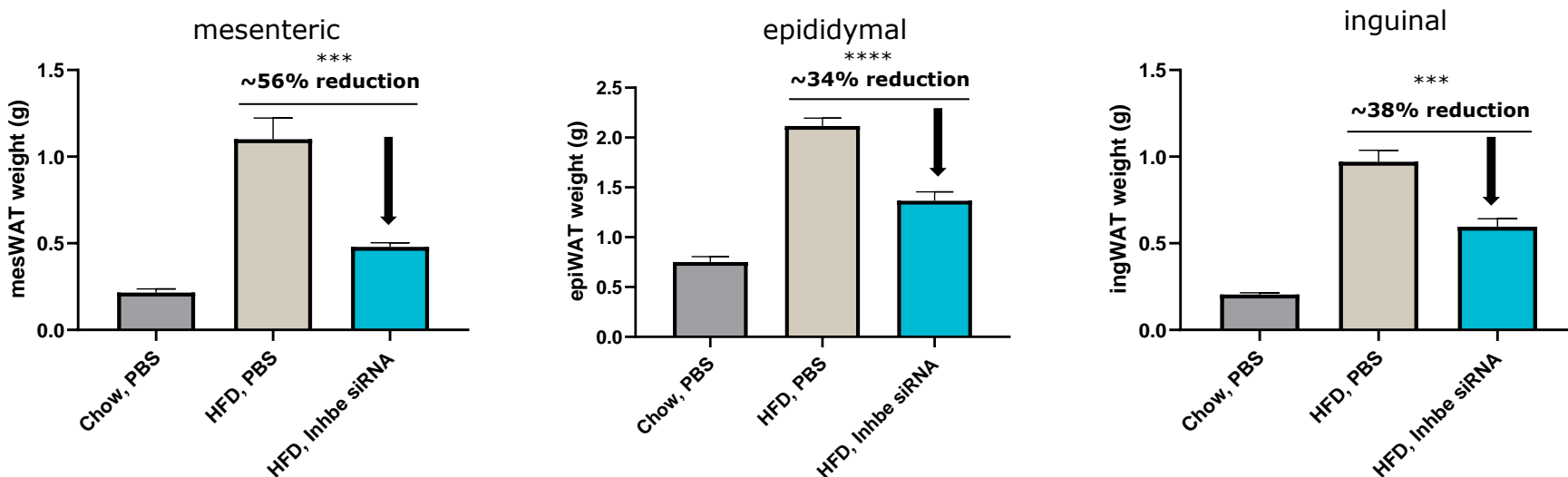
Similar effect seen in semaglutide preclinical studies



# INHBE reduction leads to significant decrease in visceral fat at 5 weeks

- INHBE knockdown in young DIO mice resulted in less fat mass across multiple types of white adipose tissue, without loss of brown fat

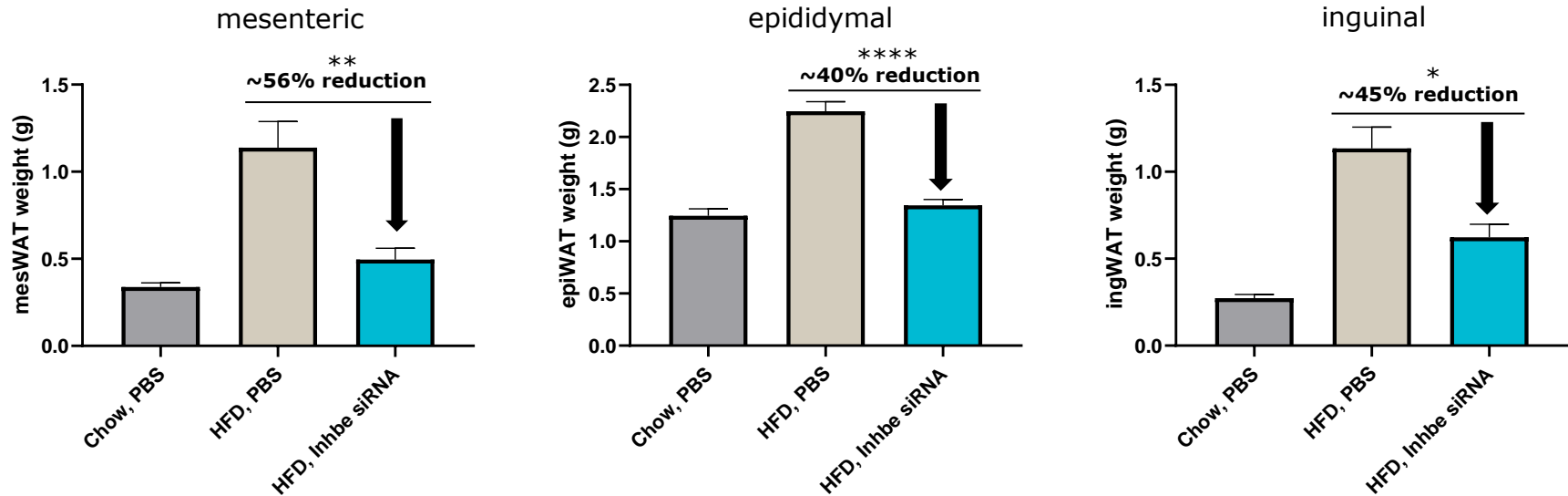
Changes in white adipose tissue after 5 weeks



# >50% reduction of INHBE mRNA recapitulates phenotype of heterozygous LoF carriers

- Subsequent 8-week study demonstrates further reduction in excess visceral fat

Changes in white adipose tissue after 8 weeks

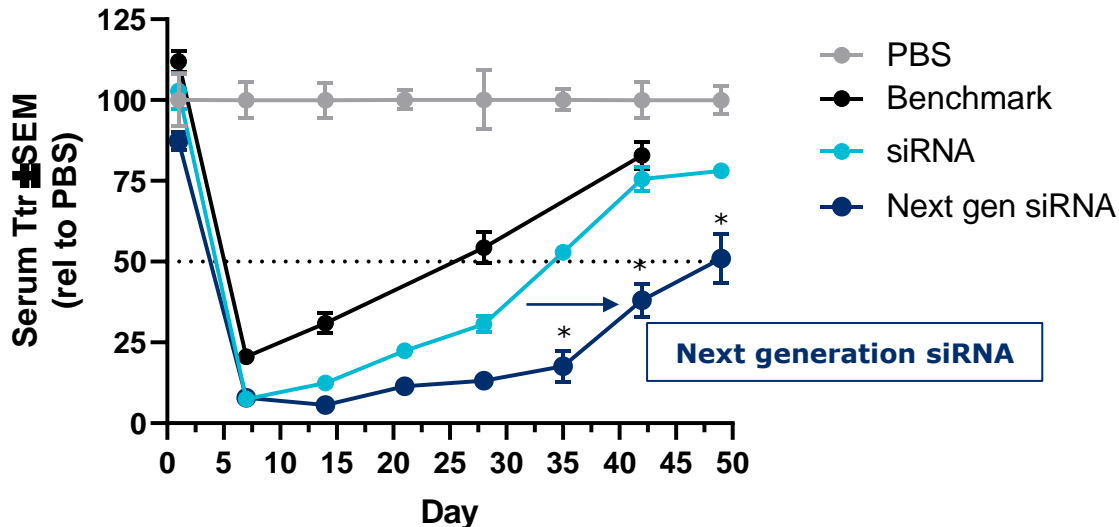


First demonstration of siRNA treatment to restore healthy phenotype



# Wave's next generation GalNAc-siRNA demonstrates best-in-class potential

Next generation siRNA results in more potent and durable knockdown of serum Ttr protein

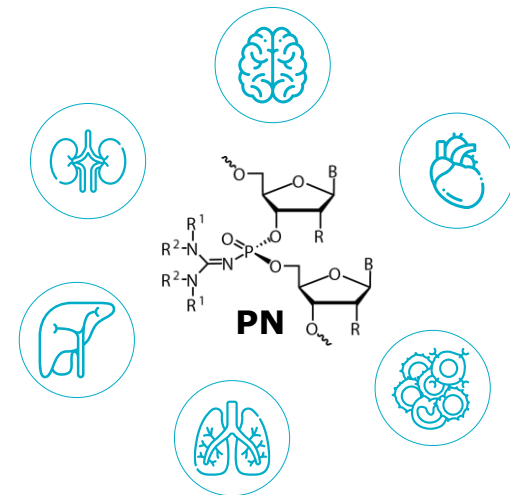


- Applying next-generation siRNA chemistry to INHBE program
- Potential for infrequent administration

**INHBE candidate for metabolic disorders, including obesity, expected in 4Q 2024**

# Wave's platform chemistry enables siRNA extra-hepatic delivery

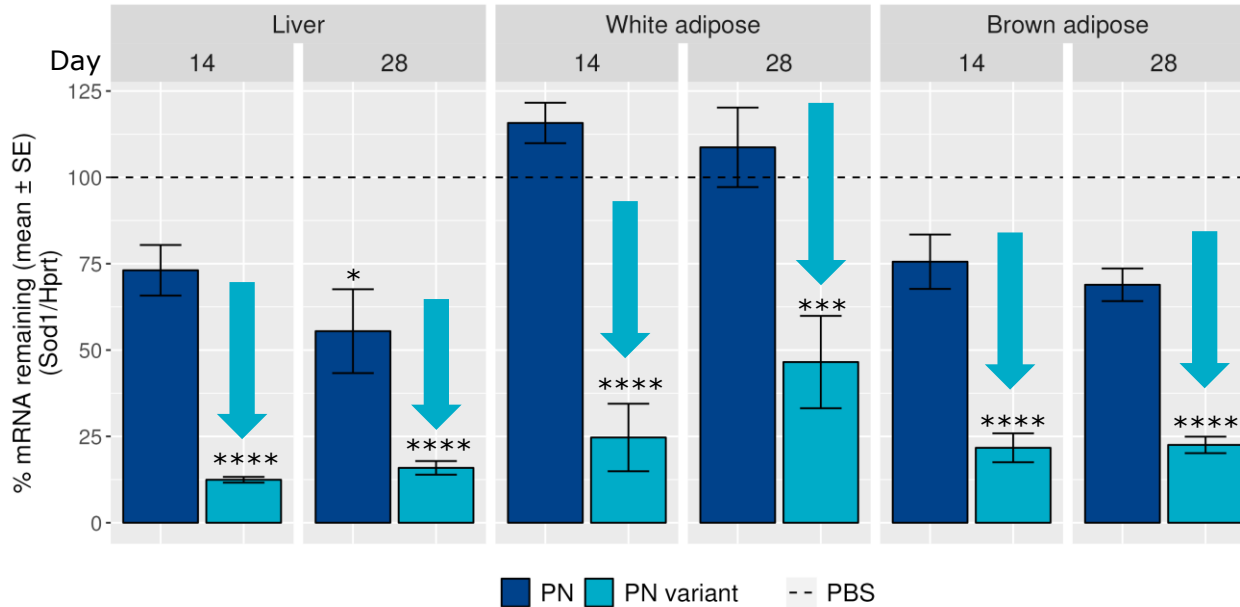
- Chemical impact
  - Introduction of neutral backbone
  - Unique structural feature of PN, specifically guanidine
  - Increased lipophilicity
  - Stereochemistry
- Extra-hepatic delivery
  - Titrating siRNA lipophilicity tunable PNs (PN variants)
  - Maintaining high Ago2 loading and intracellular trafficking
  - Titrating plasma protein binding
  - Altered delivery, enhanced potency and durability in various tissues



**PN can tune extra-hepatic delivery of siRNA using rational design, including placement, number of modifications and PN variants**

# Tunable PN variants enhance potency and alter extra-hepatic delivery of non-GalNAc siRNAs

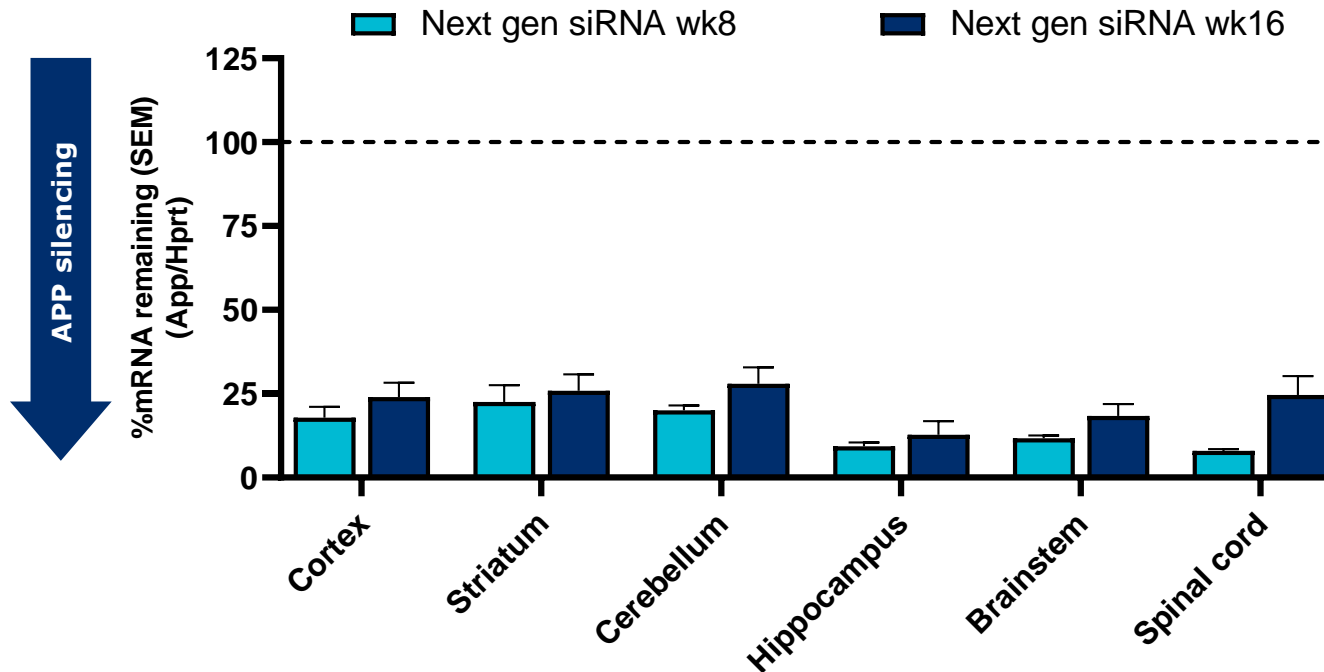
## Non-GalNAc siRNA with PN variants improve silencing in liver and adipose tissue 14 and 28 days post single dose



- Reaching adipose tissue in addition to liver with siRNA is important for certain metabolic disorders
- PN variants also enhanced siRNA silencing in muscle tissue, including heart and diaphragm

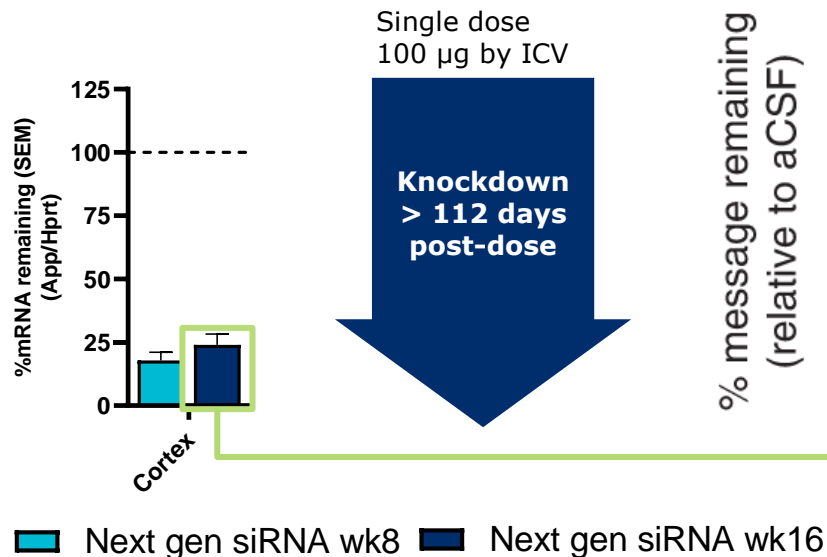
# Single dose of next generation siRNA delivers broad, potent and durable CNS target engagement

Sustained APP knockdown of at least 75% throughout the 16-week study *in vivo* in mice

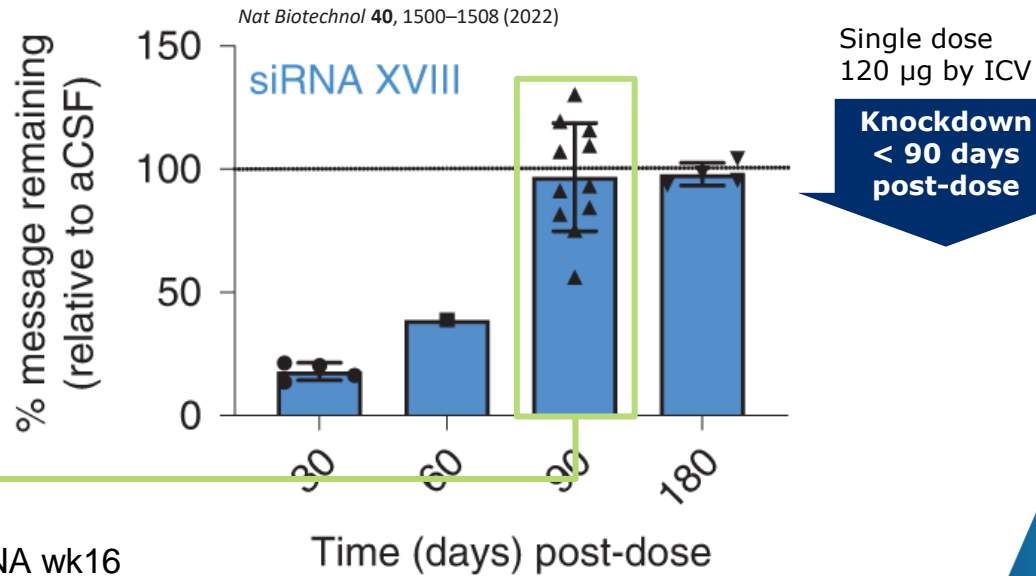


# Wave siRNA demonstrates more potent and durable silencing as compared to published state-of-the-art

## Wave (APP – Cortex)

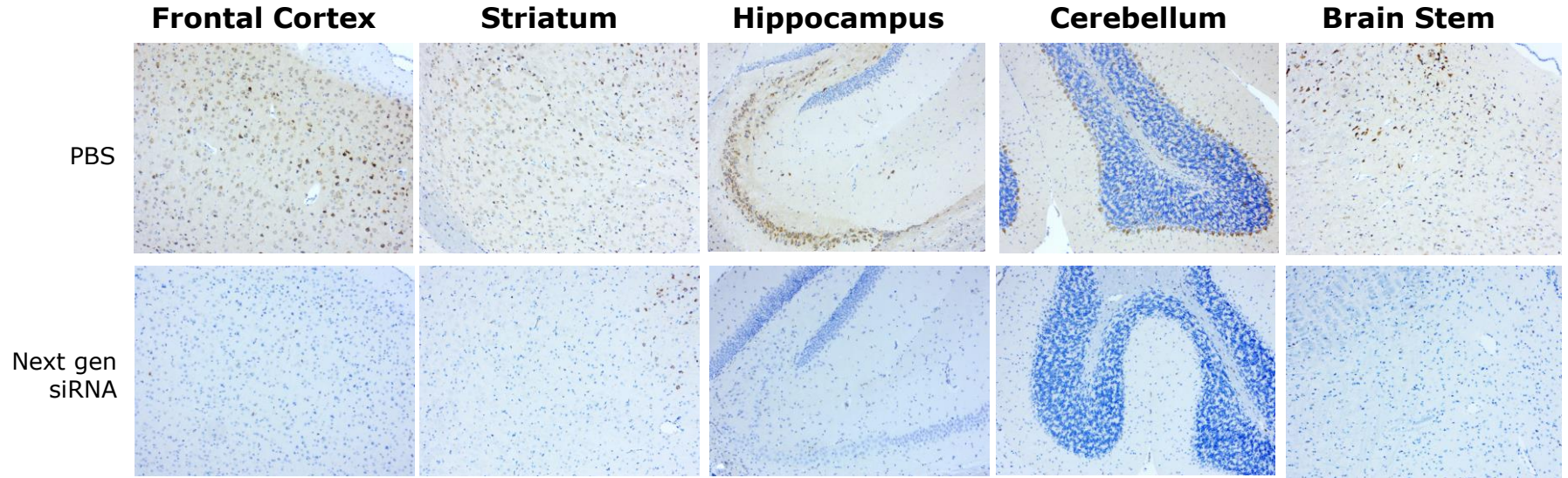


## Alnylam (APP – Cortex)



# Robust target engagement translates to substantial App protein reduction across brain regions

Reductions observed 8-weeks post single-dose



# First siRNA clinical candidate (INHBE) with proprietary chemistry expected in 4Q 2024

- **INHBE GalNAc-siRNA program is driven by clinical genetics, with potential to be next-generation therapeutic for obesity**
  - ≥50% silencing of INHBE is expected to improve metabolic health
  - INHBE siRNA silencing above therapeutic threshold restores healthy phenotype, with 16% lower body weight, as well as reduction of visceral fat to the level of lean-animals
- **Next generation GalNAc-siRNA formats are best-in-class and being applied to INHBE program**
- **Wave's platform chemistry enables extra-hepatic delivery for other non-hepatic targets**
  - PN-variants on non-GalNAc siRNA enhance silencing in multiple tissues, including liver, adipose tissue and muscle.
  - Single dose of next generation siRNA delivers broad, potent (>75%) and durable CNS target engagement



# AIMers: Editing to Upregulate

Chandra Vargeese, PhD  
*Chief Technology Officer*

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# First-generation AIMER designs published in *Nature Biotechnology*



## Endogenous ADAR-mediated RNA editing in non-human primates using stereorepchemically modified oligonucleotides

Prashant Monian<sup>1,2</sup>, Chikdu Shivallia<sup>1,2</sup>, Genliang Lu<sup>1</sup>, Mamoru Shimizu<sup>1</sup>, David Boulay<sup>1</sup>, Karley Bussow<sup>1</sup>, Michael Byrne<sup>1</sup>, Adam Bezigian<sup>1</sup>, Arindam Chatterjee<sup>1</sup>, David Chew<sup>1</sup>, Jigar Desai<sup>1</sup>, Frank Favalaro<sup>1</sup>, Jack Godfrey<sup>1</sup>, Andrew Hoss<sup>1</sup>, Naoki Iwamoto<sup>1</sup>, Tomomi Kawamoto<sup>1</sup>, Jayakanthan Kumarasamy<sup>1</sup>, Anthony Lamattina<sup>1</sup>, Amber Lindsey<sup>1</sup>, Fargjium Liu<sup>1</sup>, Richard Looby<sup>1</sup>, Subramanian Marappan<sup>1</sup>, Jake Metterville<sup>1</sup>, Ronelle Murphy<sup>1</sup>, Jeff Rossi<sup>1</sup>, Tom Fu<sup>1</sup>, Bijay Bhattacharjya<sup>1</sup>, Stephany Standley<sup>1</sup>, Snehasita Tripathi<sup>1</sup>, Halin Yang<sup>1</sup>, Yuan Yin<sup>1</sup>, Hui Yu<sup>1</sup>, Cong Zhou<sup>1</sup>, Luciano H. Apponi<sup>1</sup>, Pachamuthu Kandasamy<sup>1</sup> and Chandra Vargese<sup>1,2</sup>✉

Technologies that recruit and direct the activity of endogenous RNA-editing enzymes to specific cellular RNAs have therapeutic potential, but translating them from cell culture into animal models has been challenging. Here we describe short, chemically modified oligonucleotides called AIMers that direct efficient and specific A-to-I editing of endogenous transcripts by endogenous adenosine deaminases acting on RNA (ADAR) enzymes, including the ubiquitously and constitutively expressed ADAR1 p110 isoform. We show that fully chemically modified AIMers with chimeric backbones containing stereorepchemically modified and nitrogen-containing linkages based on phosphoryl guanidine enhanced potency and editing efficiency 100-fold compared with those with uniformly phosphorothioate-modified backbones *in vitro*. *In vivo*, AIMers targeted to hepatocytes with N-acetylglucosamine achieve up to 50% editing with no bystander editing of the endogenous *CTB* transcript in non-human primate liver, with editing persisting for at least one month. These results support further investigation of the therapeutic potential of stereorep AIMers.

Recruiting endogenous RNA-editing enzymes using chemically modified oligonucleotides holds promise for treating human disease. The most common mutation in human genes is transition from cytosine (C) to thymine (T), and CpG dinucleotides are well established hot spots for disease-causing mutations. The ADAR family of enzymes catalyze adenosine (A) to inosine (I) changes in the transcriptome<sup>1,2</sup>. Inosine is read as guanine (G) by the translational machinery<sup>3,4</sup>. ADAR-mediated RNA editing has the potential to revert these disease-causing transitions at the RNA level. The potential scope for application of A-to-I editing is large, including modulation of polar or charged amino acids, stop codons or RNA regulatory sequences<sup>5,6</sup>, eliciting disease-modified outcomes (for example, metastasis protein expression or function)<sup>7,8</sup>.

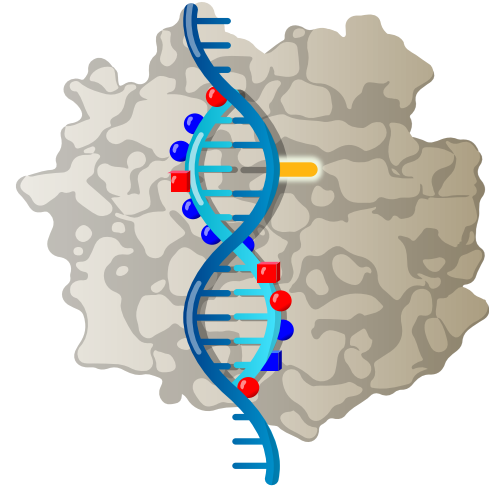
Chemical modifications are known to confer drug-like properties to oligonucleotides. We set out to determine whether control over backbone chemistry and stereochemistry and other chemical modifications to an oligonucleotide (Fig. 1 and Supplementary Note 1) can be optimized to elicit sequence-specific A-to-I RNA editing with endogenous ADAR enzymes. As therapeutic, reversible RNA editing with oligonucleotides may represent a safer option than those that edit genomic DNA. Early technologies designed to elicit RNA editing *in vitro* required an exogenous enzyme and an oligonucleotide<sup>9–11</sup>. These approaches led to overexpression of editing enzyme and substantial off-target editing<sup>12–14</sup>. Recent advances have overcome the need for exogenous enzymes *in vivo*<sup>15</sup>, but they still use long oligonucleotides that require auxiliary delivery

vehicles, such as viral vectors or lipid nanoparticles, for application beyond cell culture. So far, these technologies have yielded minimal editing *in vivo*<sup>16</sup>.

Leveraging our oligonucleotide chemistry platform, we developed relatively short oligonucleotides that direct A-to-I RNA editing with high efficiency using endogenous ADAR enzymes. These oligonucleotides, called AIMers, are short and fully chemically modified with stereorep phosphorothioate (PS) and nitrogen-containing (PN) linkages based on phosphoryl guanidine. *In vitro*, they enhanced potency and A-to-I editing efficiency compared to uniformly PS-modified AIMers, and *in vivo*, N-acetylglucosamine (GalNAc)-modified AIMers achieved up to 50% editing with no bystander editing in non-human primate (NHP) liver that persisted for at least 1 month.

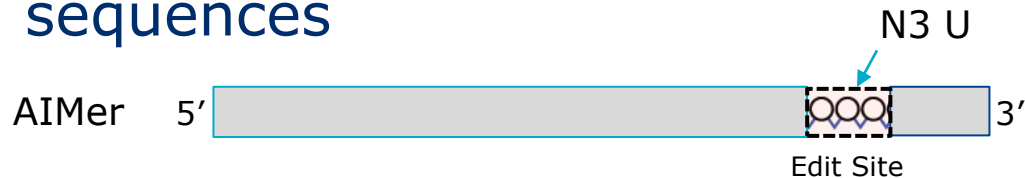
**Results**  
AIMers support RNA editing. To evaluate RNA-editing efficiency in mammalian cells, we created a luciferase reporter with genes from *Gaussia* (*Glc*) and *Cyprinus* (*Clac*). In the absence of editing, only *Glc* is expressed, whereas A-to-I editing permits expression of *Clac*, providing a measure of RNA-editing efficiency and protein expression (Extended Data Fig. 1a). AIMers were designed to mimic naturally occurring double-stranded RNA-ADAR substrates, as in the *Clac2* transcript<sup>17,18</sup> (Extended Data Fig. 1b). To benchmark RNA editing, we transfected 293T cells with the reporter and exogenous ADAR enzyme in the presence or absence

- Foundational AIMER structure-activity-relationships
- *In vitro-in vivo* translation (NHPs)
- Specificity *in vitro* & *in vivo* (NHPs)
- GalNAc conjugation

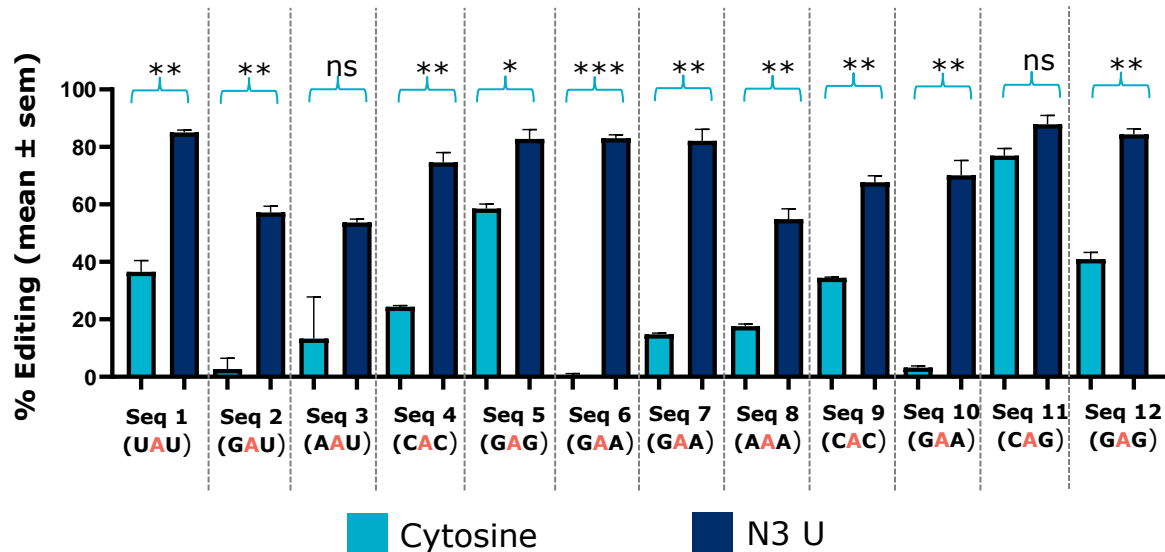


<sup>1</sup>Wave Life Sciences, Cambridge, MA, USA. <sup>2</sup>These authors contributed equally: Prashant Monian, Chikdu Shivallia. ✉e-mail: cvargese@wavelife.com

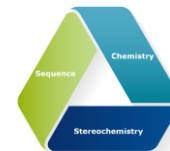
# Proprietary base modifications increase editing across edit region sequences



## Proprietary base modification (N3 U) increases *UGP2* RNA editing across sequences *in vitro*



- N3 U: example of proprietary base modifications
- N3 U consistently improves RNA editing levels, including across sequences



# Innovating on applications of ADAR editing

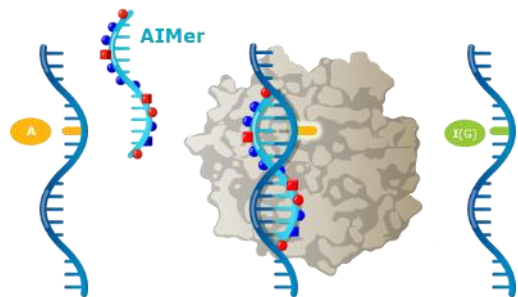
## Correct G-to-A driver mutations with AIMers

## Modulate protein interactions with AIMers



Restore or correct protein function

**WVE-006**  
(GalNAc AIMER)  
AATD



*Achieved POC*

- Modulate protein-protein interaction**
- Upregulate expression**
- Modify function
- Post-translational modification
- Alter folding or processing



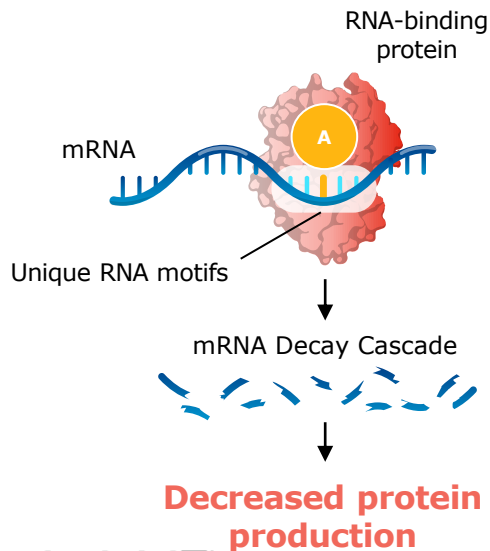
# RNA is highly regulated, creating ample opportunity to intervene with AIMers to alter protein expression



RNA binding proteins recognize sequence motifs to regulate mRNA stability



## Attenuated Gene Expression



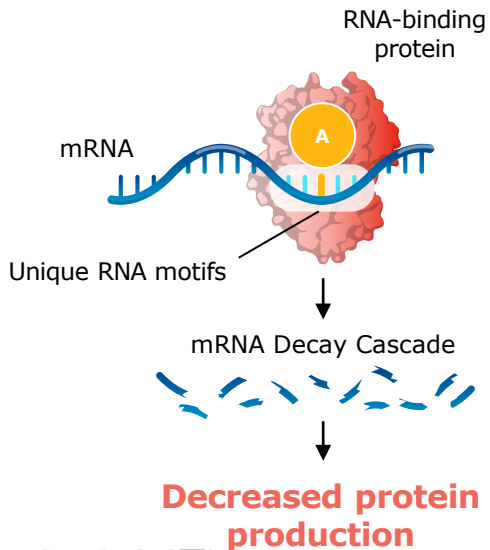


# Using catalytically efficient AIMers, mRNA can be edited and durably stabilized

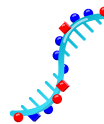
RNA binding proteins recognize sequence motifs to regulate mRNA stability



**Attenuated  
Gene Expression**



**“Dialed up”  
Gene Expression**



**Catalytically Efficient Aimer**

Aimer **edits** and **durably stabilizes** mRNA



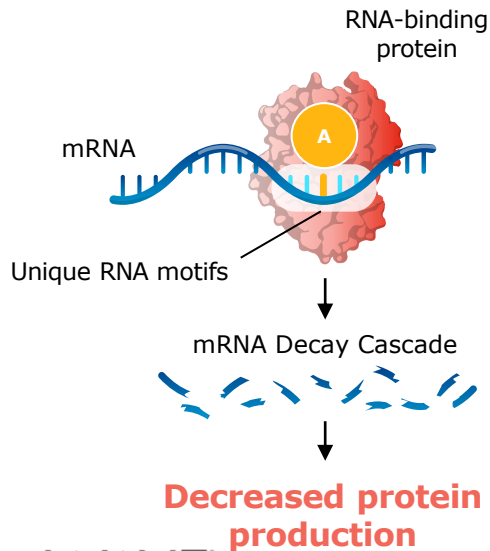


# Upregulation: AIMers can edit RNA motifs to restore or upregulate gene expression

RNA binding proteins recognize sequence motifs to regulate mRNA stability



**Attenuated Gene Expression**

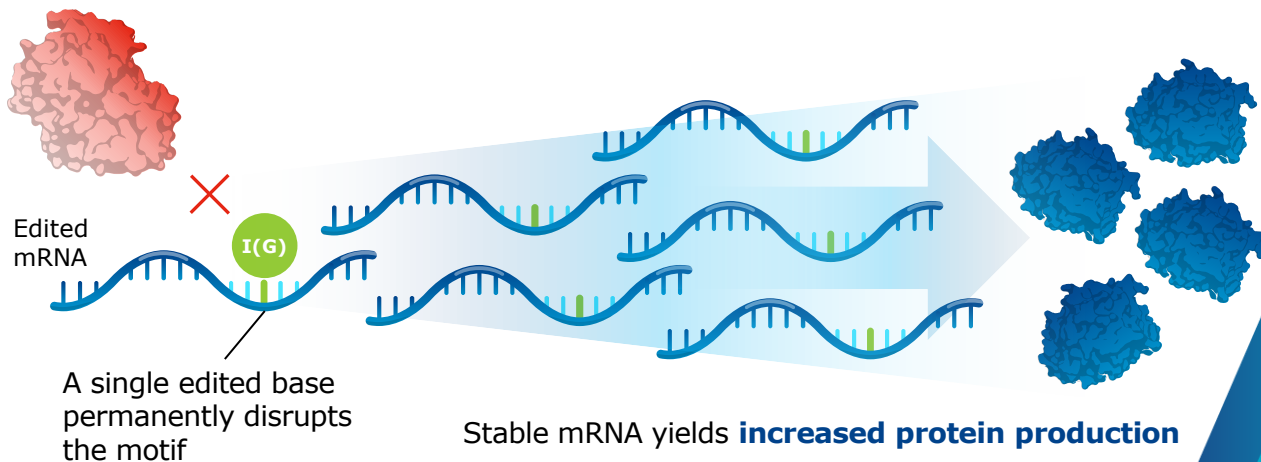


**"Dialed up" Gene Expression**



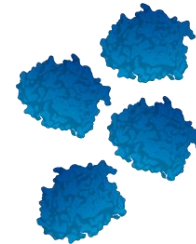
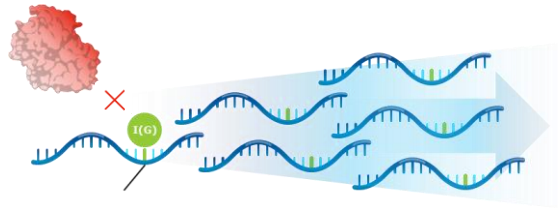
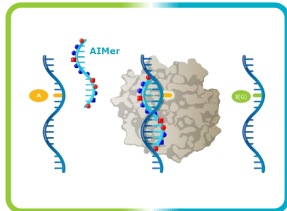
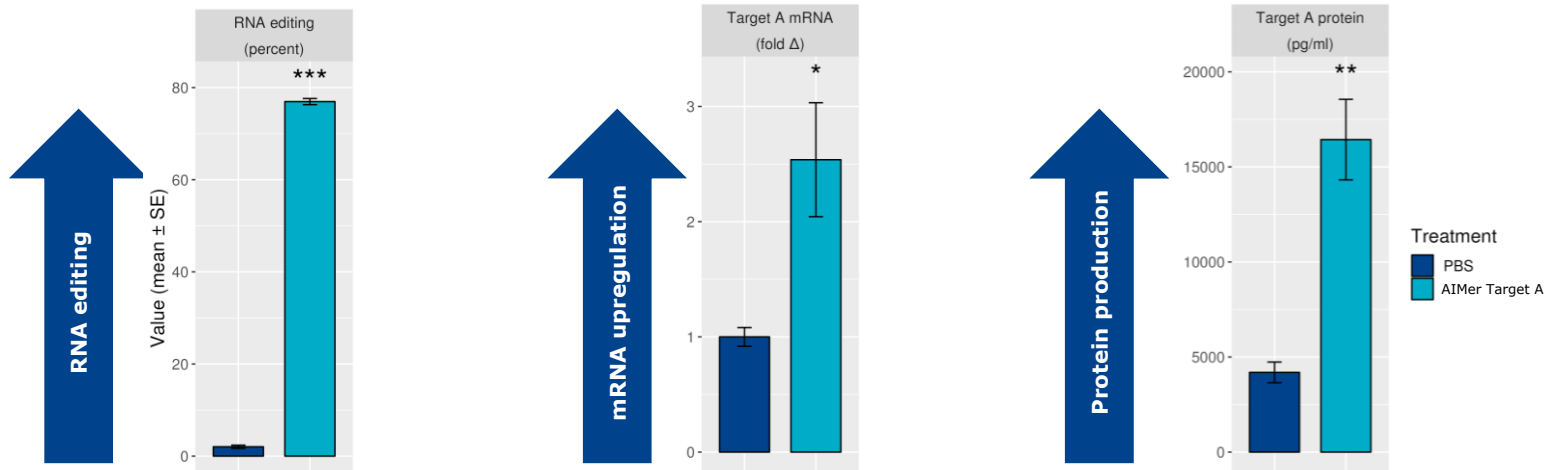
**Catalytically Efficient AIMer**

AIMer **edits** and **durably stabilizes** mRNA



# Proof-of-concept: Editing RNA motifs to upregulate mRNA and increase protein

RNA editing, mRNA upregulation and protein expression *in vivo* for an undisclosed Target A





## Mapping the “Edit-verse”

Kenneth Longo, PhD  
*Vice President,  
Data Science*

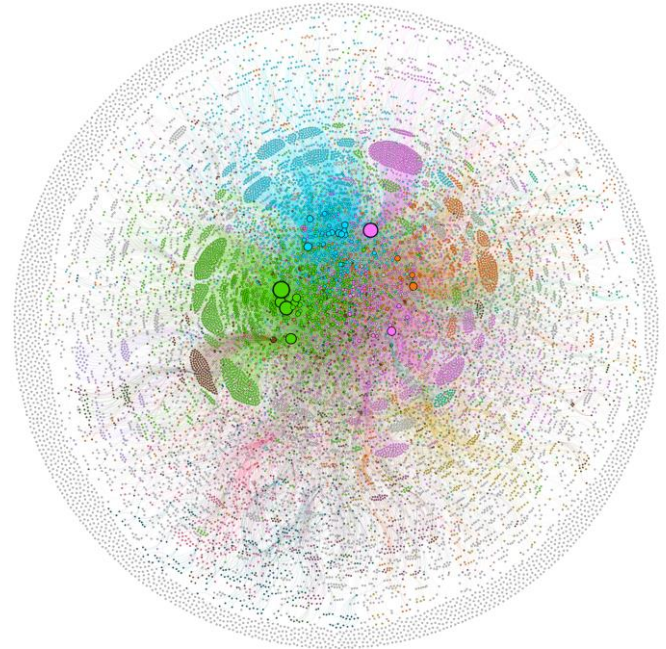
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# Mapping the RNA editing target universe

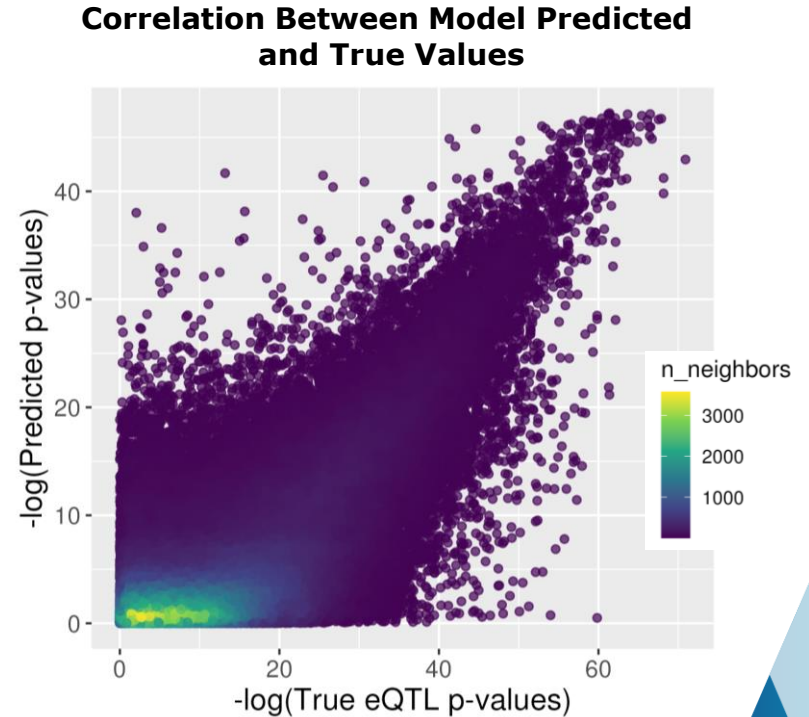
- The editable gene-disease network, **“The Edit-Verse”**, is enormous and includes coding and non-coding regions of transcripts
- The **upregulation target universe** is particularly interesting because many diseases are associated with reduced protein expression:
  - Haploinsufficient and hypomorphic variants
  - Regulatory variants
- Upregulation offers the potential to address multiple pathogenic mutations **with a single therapy**

**Gene-Disease Network**



# Wave's deep learning model predicts novel edit sites that impact transcriptional regulation

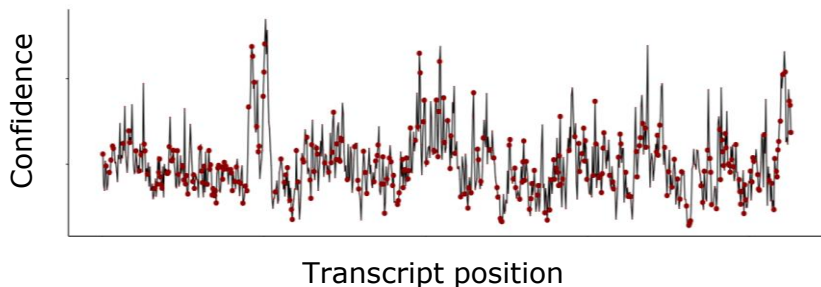
- Proprietary model constructed using large expression quantitative trait loci (eQTL) databases that can predict the impact of editing on gene expression
- Model achieved good predictive accuracy on known eQTLs
- Results include long list of *novel* eQTL sites where an A-to-G edit, never-before observed in nature, confidently predicts changes in transcript levels for >50% of proteome
- Ongoing model development is expected to expand Edit-verse further



# Identify AIMer-mediated upregulation opportunities in disease sub-networks

- For instance, we can zoom into network for the hyperlipidemia and energy intake GWAS phenotypes, which contains 96 genes and disease-pathway associations
- We can then make editing predictions for any in-network gene of interest

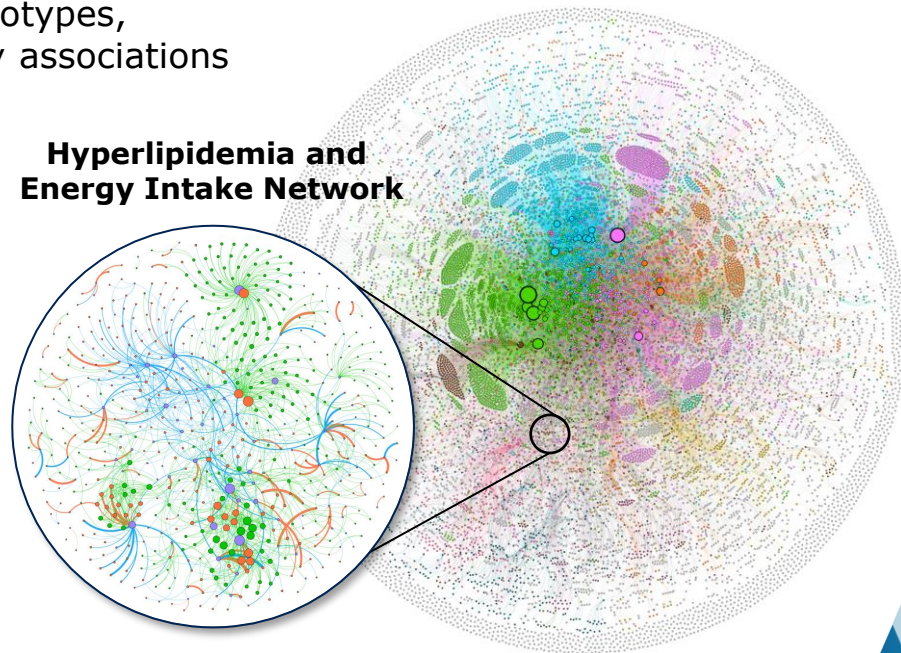
## ADAR Amenable Sites Predicted to Impact Half-Life



● Potential AIMer-targetable adenosines

## Gene-Disease Network

## Hyperlipidemia and Energy Intake Network





## Mining the “Edit-verse”

Ginnie Yang, PhD  
*Senior Vice President,  
Translational Medicine*

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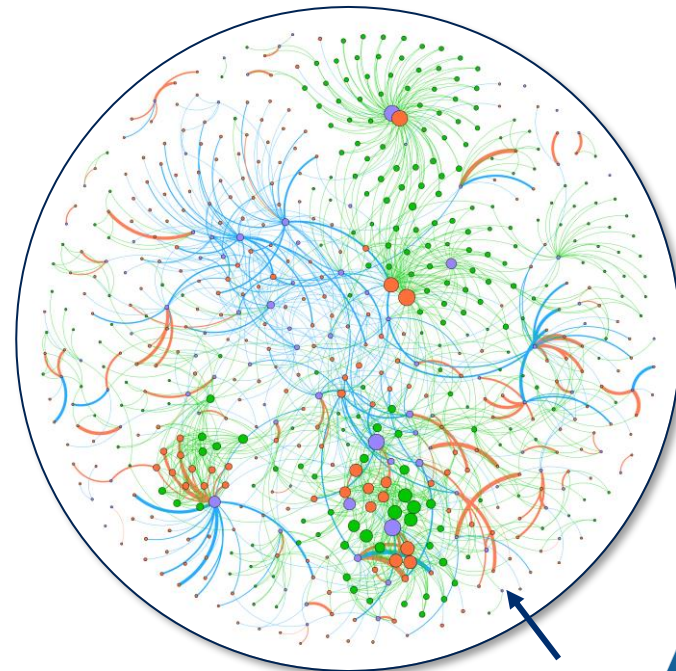
# Edit-verse subnetwork reveals “Target A”: Metabolic syndrome target uniquely suited for AIMer upregulation



## Target A

- Liver target for upregulation, non-incretin therapy
- Strongly implicated in metabolic disease, with indirect causation in familial disorders
- Few therapies today provide weight loss in this specific patient population
- Estimate 90 million potential patients in the US and Europe with metabolic syndrome and obesity
- Serum protein levels and biomarkers available to assess target engagement

## Hyperlipidemia and Energy Intake Network



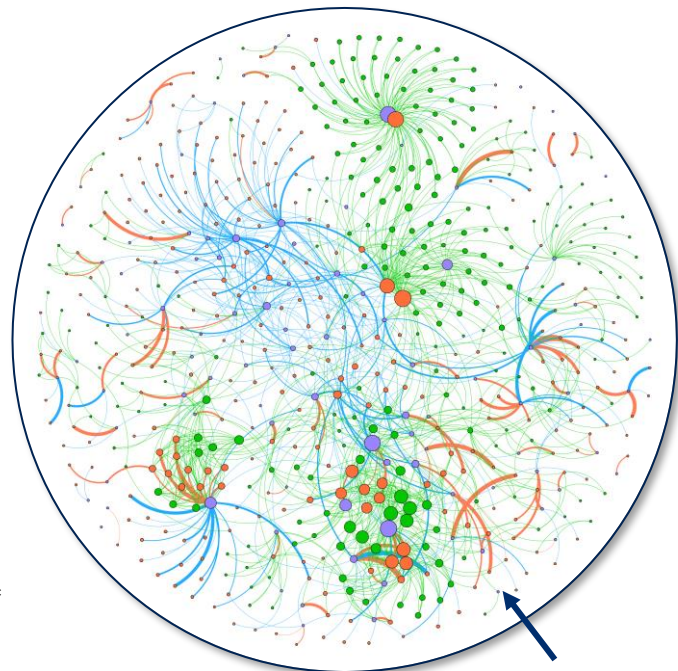
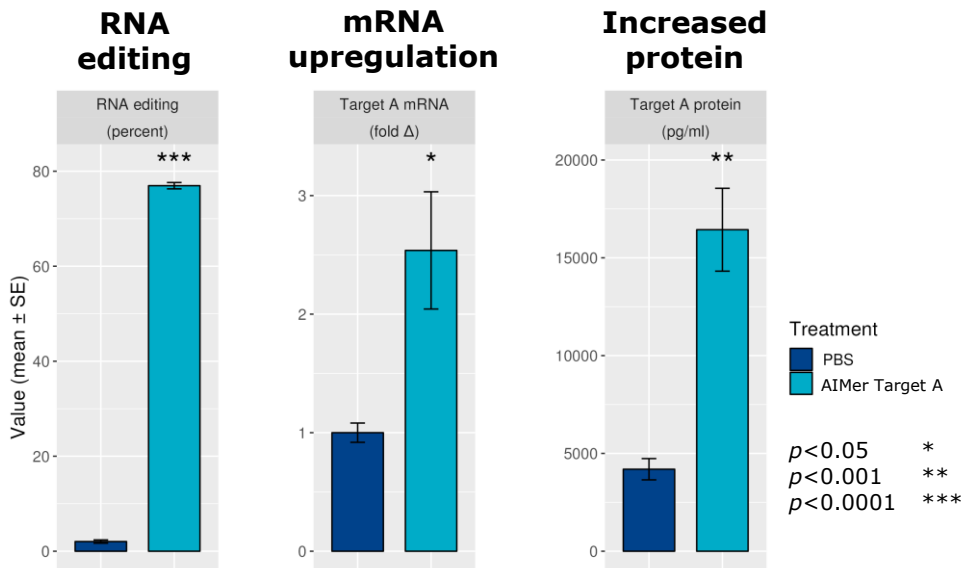
Target A



# First preclinical *in vivo* PoC: upregulating endogenous protein to restore healthy metabolic phenotype

>75% RNA editing led to >2-fold increase of mRNA, and similar degree of protein upregulation *in vivo* with GalNAc-AIMER in young DIO mice

## Hyperlipidemia and Energy Intake Network



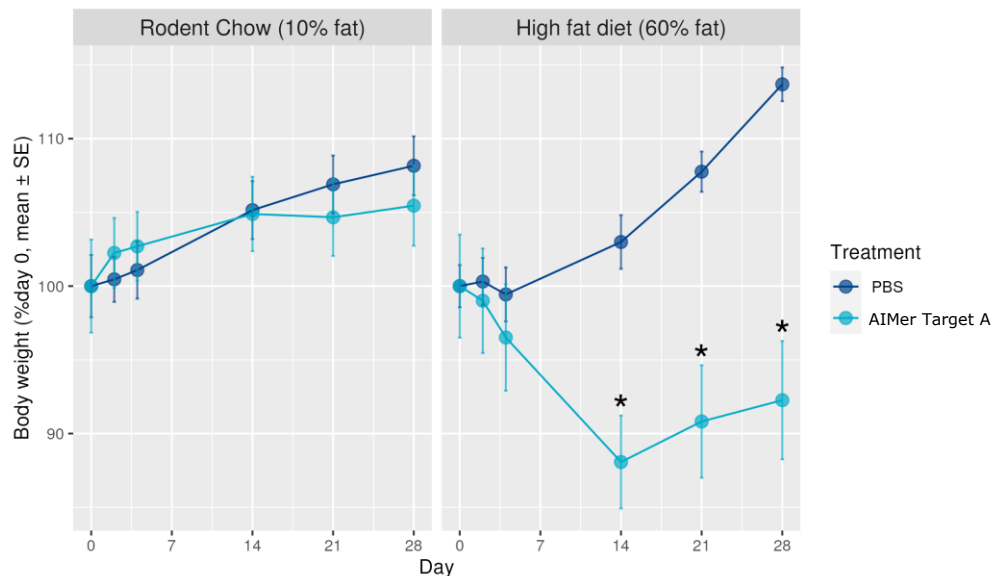
Target A



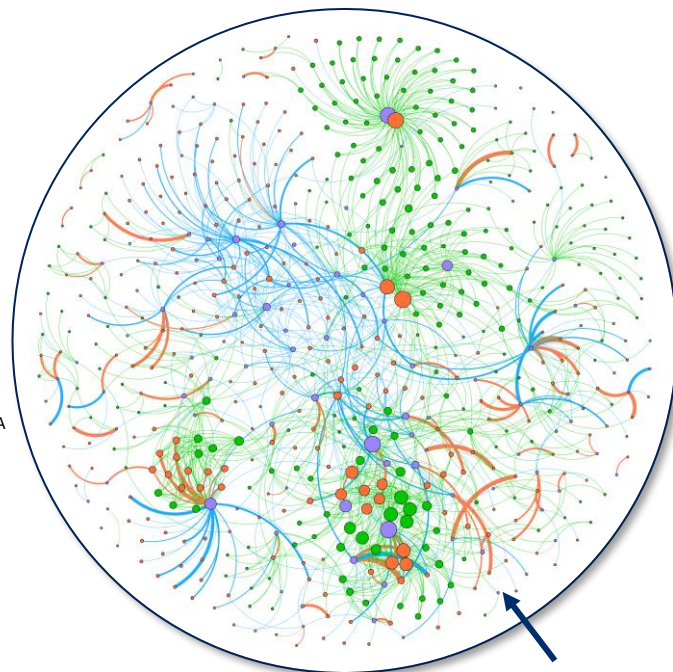
# Substantial upregulation of protein induces weight loss

- ~3-fold upregulation of Target A protein with GalNAc-AIMER led to weight reduction in DIO mice

## Significant Weight Loss



## Hyperlipidemia and Energy Intake Network



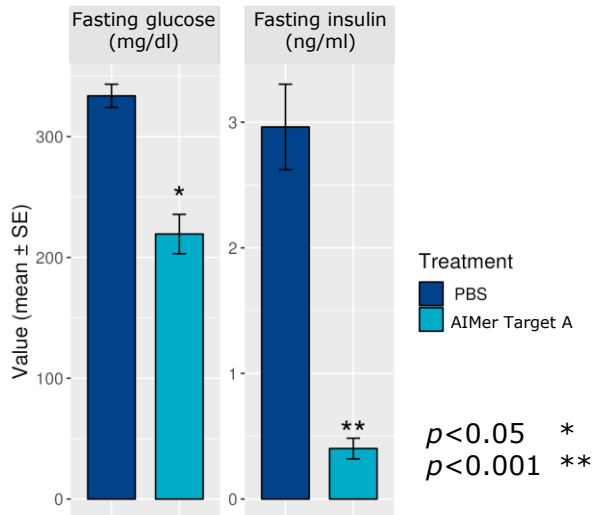
Target A

# Upregulation of Target A protein improves insulin sensitivity

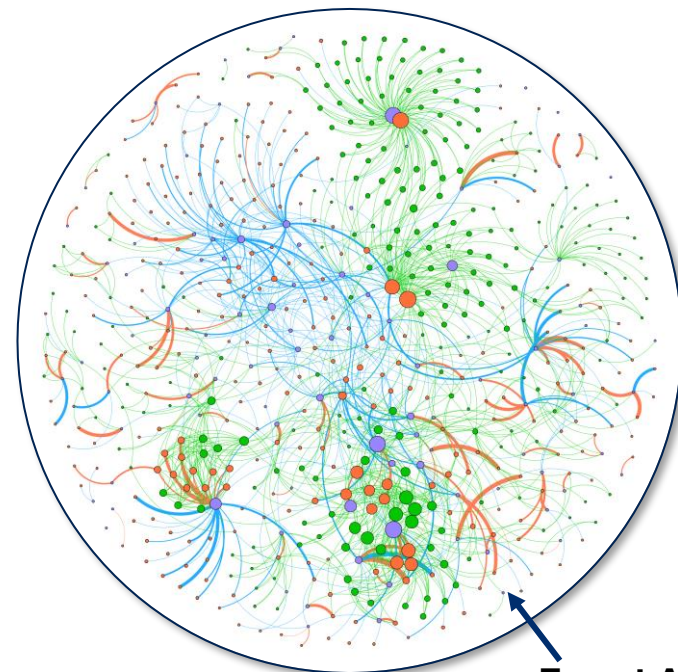


- ~3-fold upregulation of Target A protein with GalNAc-AIMER led to improved insulin sensitivity in DIO mice

## Improved Insulin Sensitivity



## Hyperlipidemia and Energy Intake Network



Target A



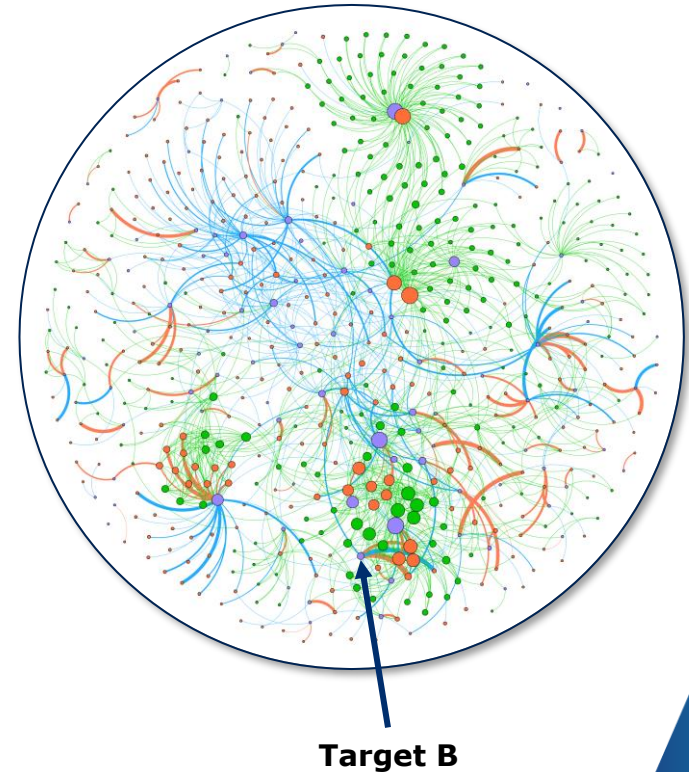
# Target B upregulation offers a first-in-class therapeutic approach for hyperlipidemia



## Target B

- Liver target for upregulation
- Hyperlipidemia; first-in-class therapeutic approach
- Estimate ~3 million target patients in US and Europe
- Serum biomarkers available to assess target engagement and efficacy
- Potential clinically meaningful benefit of >2 fold upregulation of target mRNA

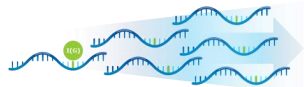
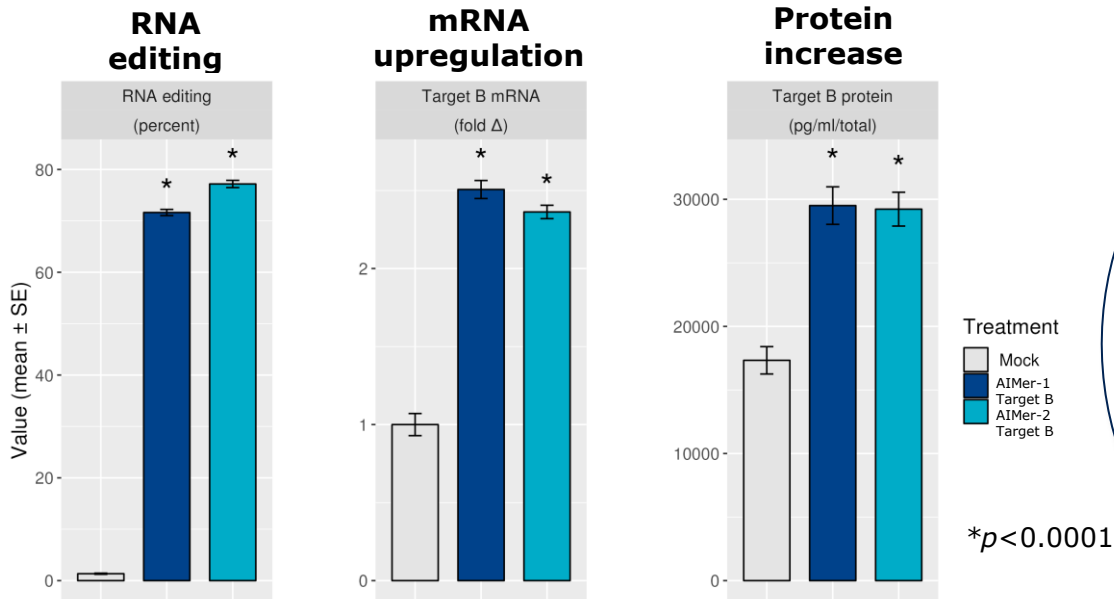
## Hyperlipidemia and Energy Intake Network



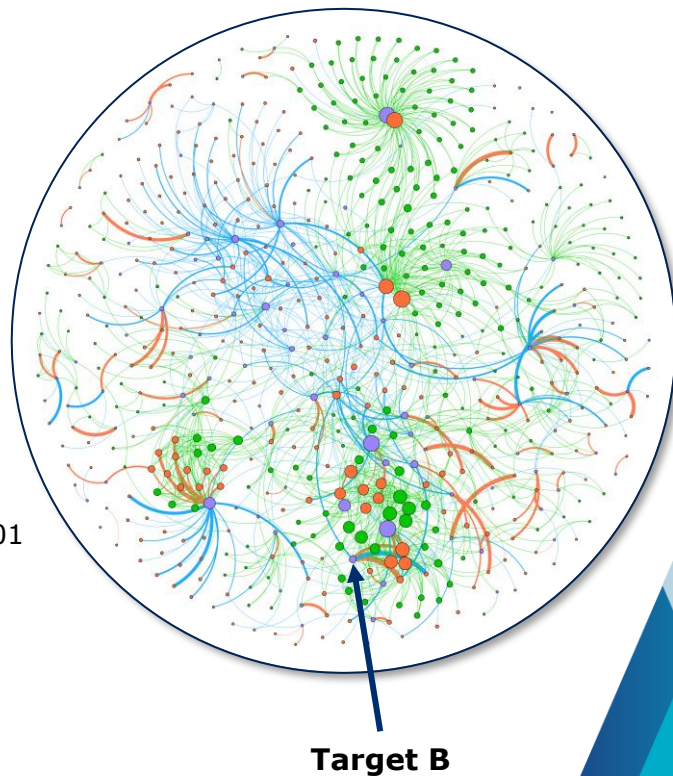


# >70% editing achieves ~2-fold upregulation with corresponding increase in protein

Primary human hepatocytes *in vitro*



## Hyperlipidemia and Energy Intake Network



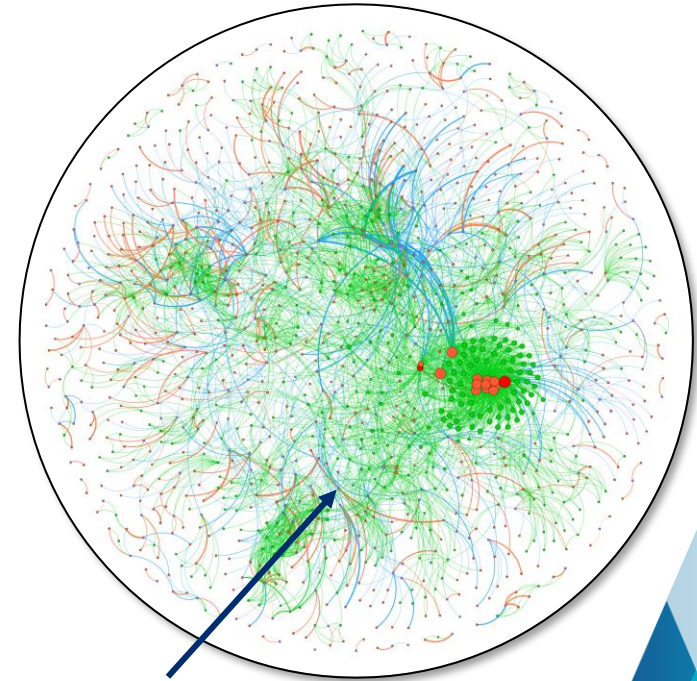
# Upregulation of liver Target X stops decline in kidney function



## Target X

- Liver target for upregulation
- Target X produces a secreted protein to treat kidney disease
- Estimate ~170K target patients in US and Europe
- Therapeutic rationale supported by genetic insights, PheWAS, and observational data
- Plasma biomarkers available to assess target engagement
- ~2-fold upregulation in secreted protein expected to be clinically meaningful

## Renal Insufficiency Network



Target X

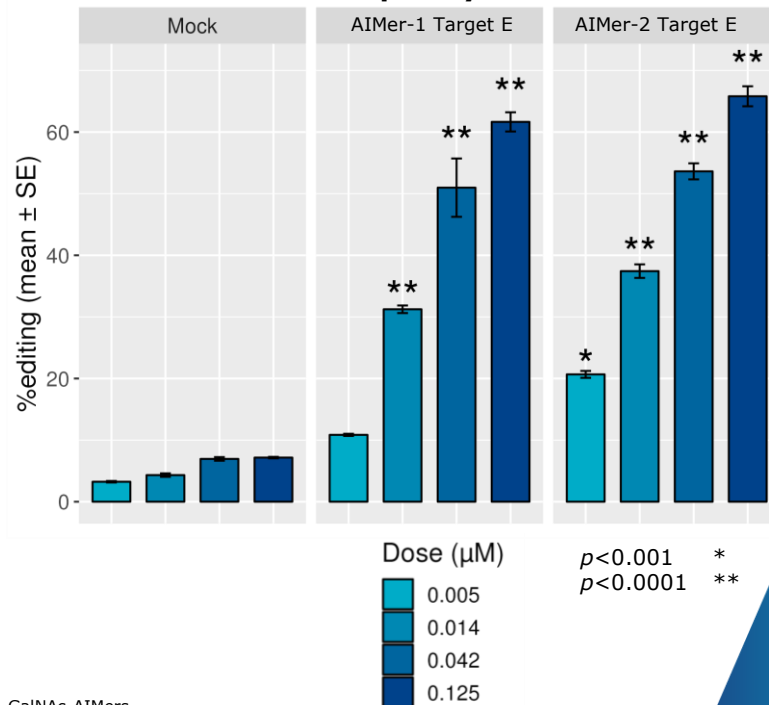


# Building on success of AATD: Target E correction restores normal metabolism in rare genetic disease

## Target E

- Liver target for correction
- Rare genetic disease
- High unmet need population not addressed with current therapeutic options
- ~17,000 patients addressable with correction approaches in US and Europe
- Fully translatable serum biomarker
- ~15-30% editing expected to deliver clinically meaningful benefit

## Proof-of-concept RNA editing in human primary hepatocytes



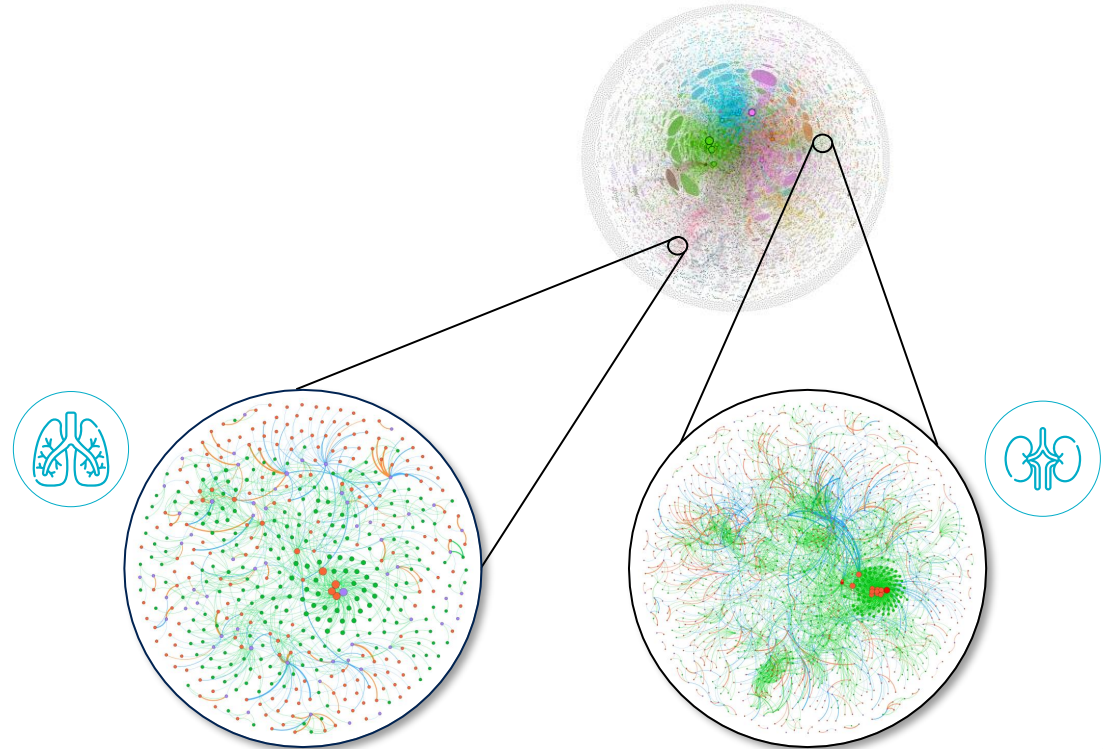
# AIMer targetable diseasomes in extra-hepatic organs

## Hepatic

- Target A
- Target B
- Target X
- Target E

## Extra-hepatic

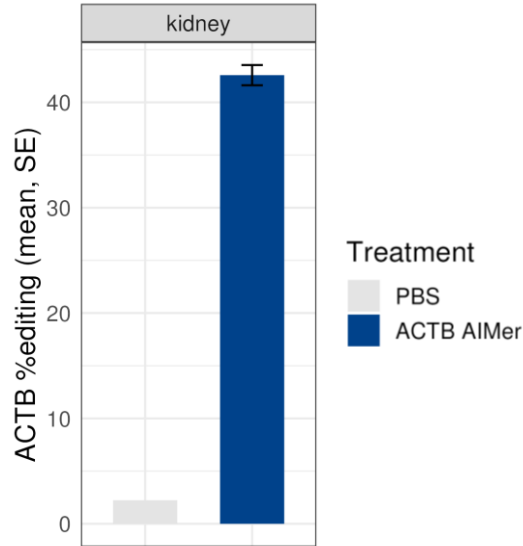
- Target F
- Target G



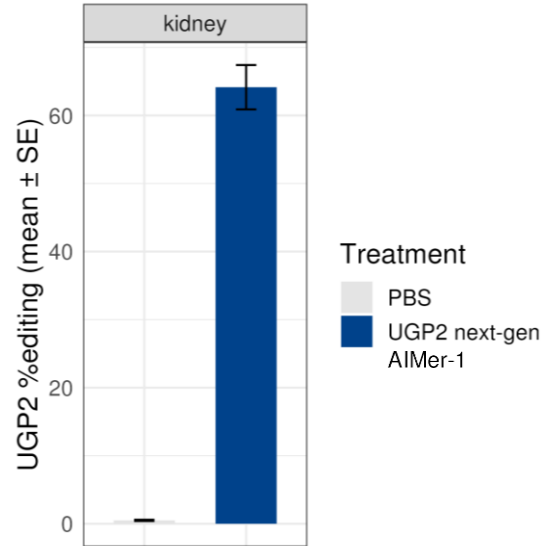
# AIMers deliver to proximal and distal convoluted tubules of kidney and achieve substantial editing



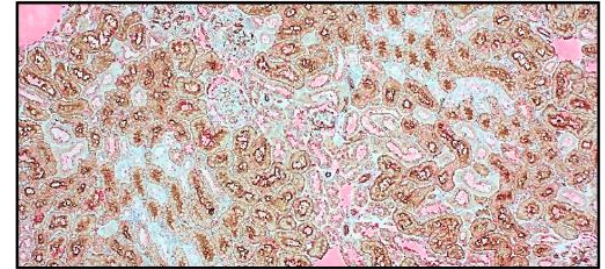
**~40% editing of ACTB in NHP  
1-week post-single dose (SC)**



**~60% editing of UGP2 in mice  
1-week post single dose (IV)**



**AIMers (red) accumulated in proximal convoluted tubules (brown) in the NHP kidney following subcutaneous administration**



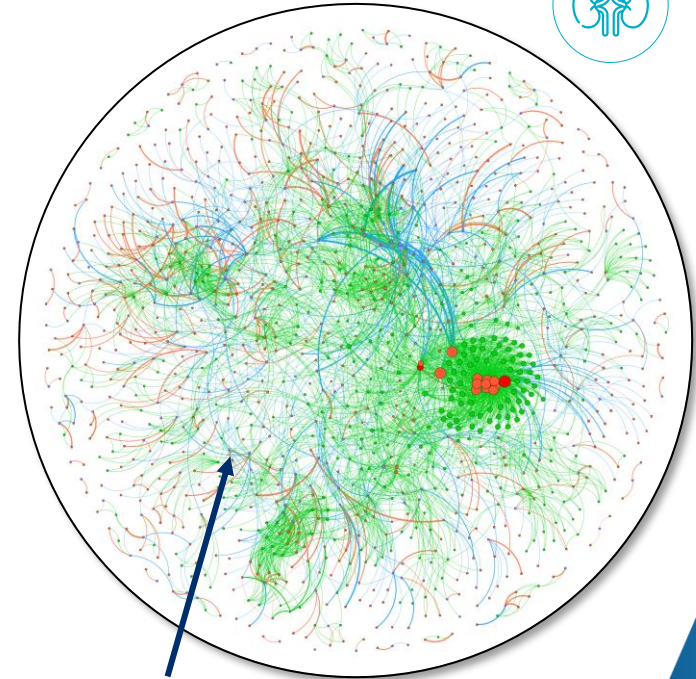
# Upregulation of Target F restores kidney function in a rare genetic kidney disease



## Target F

- Kidney target for upregulation
- Rare genetic kidney disease that leads to ESRD and need for dialysis / transplantation; High unmet need with few treatment options currently available
- ~85K patients in US and Europe addressable with upregulation approach
- Urinary biomarkers available to assess upregulation
- Clinically meaningful benefit may be achieved with 2-fold upregulation

## Renal Insufficiency Network

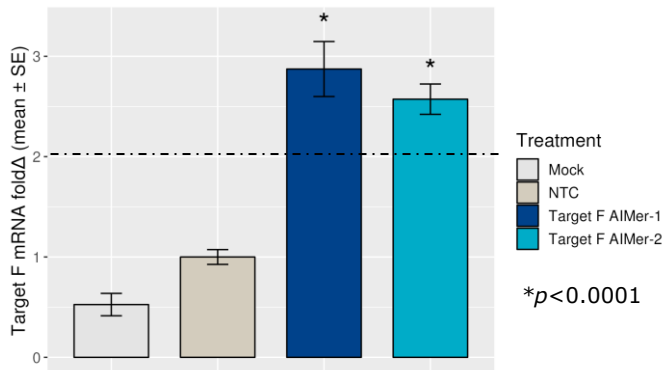


Target F

# Achieved >2-fold upregulation of Target F mRNA *in vitro* with RNA editing



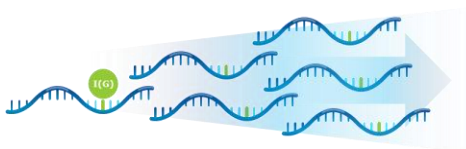
## Upregulation of Target F mRNA in Human kidney tubular epithelial cells



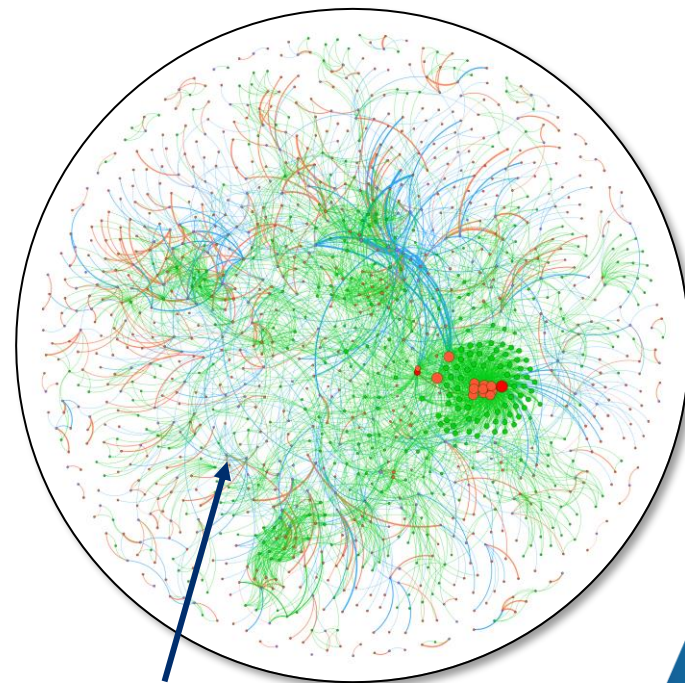
Treatment

- Mock
- NTC
- Target F Aimer-1
- Target F Aimer-2

\* $p < 0.0001$



## Renal Insufficiency Network



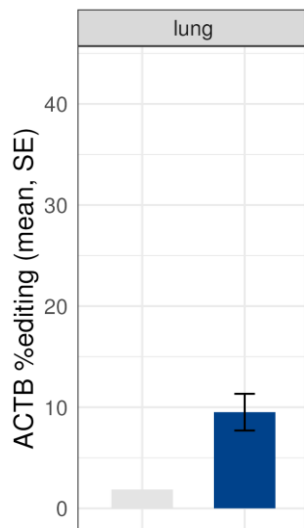
Target F



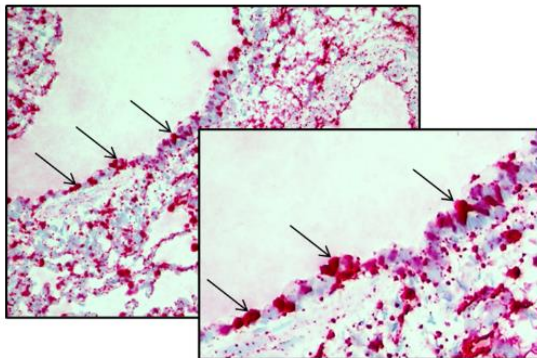
# Proprietary AIMer modifications enhance delivery to lung tissue and achieve significant editing *in vivo*



~10% editing of ACTB in NHP  
1-week post-single dose (SC)

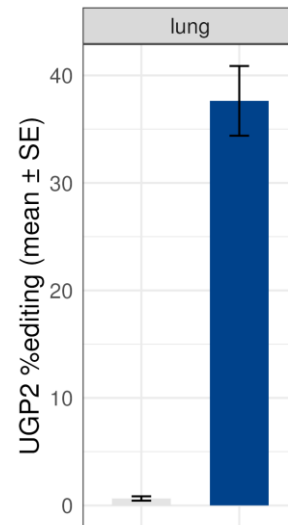


ACTB AIMer (red) deliver to bronchial epithelial cells (arrows)



Treatment  
■ PBS  
■ ACTB AIMer

>35% editing of UGP2 in mice  
1-week post single dose (IV)



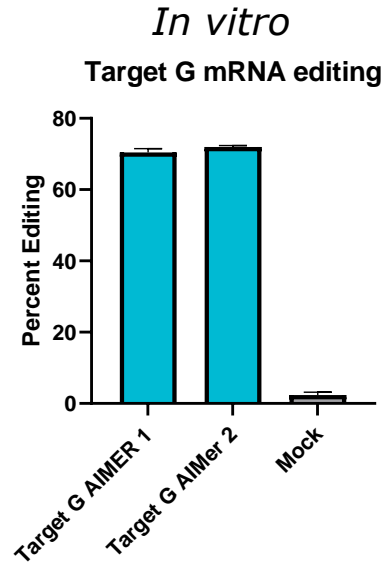
Treatment  
■ PBS  
■ UGP2 next-gen AIMer-2

# Correction of Target G mutation restores protein function in patients with a genetic lung disease

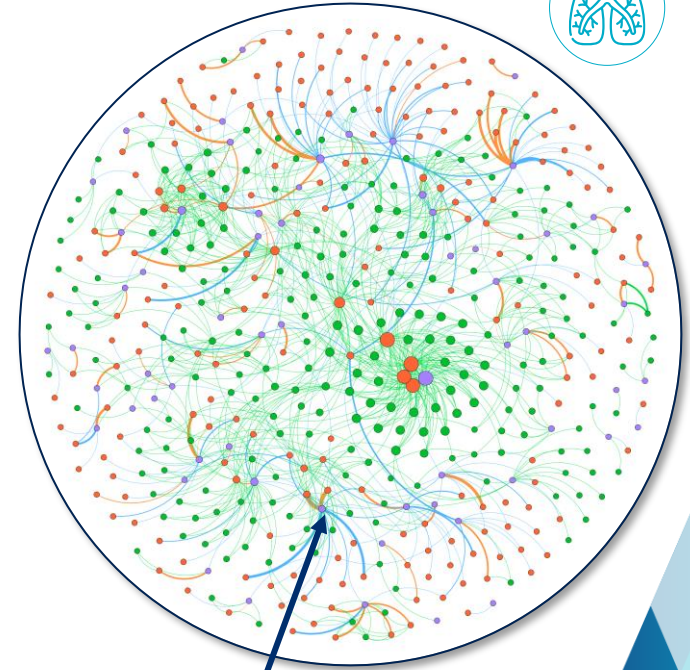


## Target G

- Lung disease target for correction
- Genetic lung disease with target patient population not addressed with available therapies
- ~5K patients amenable to correction approaches in US and Europe
- Clinically meaningful benefit expected with 20% correction
- Established clinical regulatory pathway



## Pulmonary Fibrosis Network



Target G

# Multiple RNA editing opportunities to build high-value pipeline beyond WVE-006

- The Edit-verse is substantial and still expanding
- Advancing work for a diverse set of undisclosed targets addressing areas of high unmet need, including both rare and prevalent diseases

## Potential to advance any combination of targets into preclinical development

	Hepatic (GalNAc-AIMers)				Extra-Hepatic (AIMers)	
	Target A	Target B	Target X	Target E	Target F	Target G
Approach	Upregulation	Upregulation	Upregulation	Correction	Upregulation	Correction
Tissue	Liver	Liver	Liver	Liver	Kidney	Lung
Therapeutic Area	Metabolic	Metabolic	Renal	Rare	Renal	Rare
Estimated Patients (US and Europe)	~90M	~3M	~170K	~17K	~85K	~5K

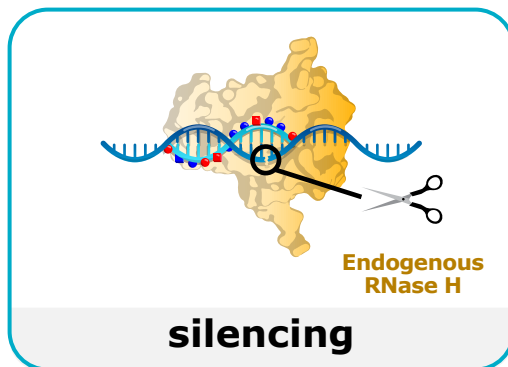
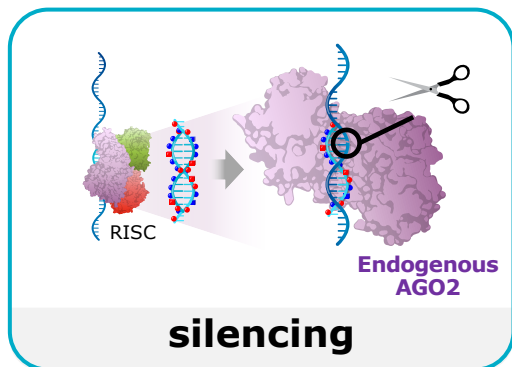
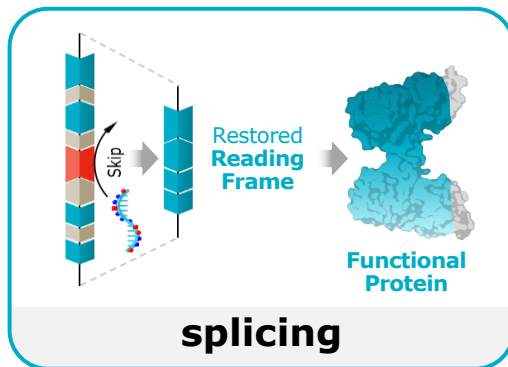
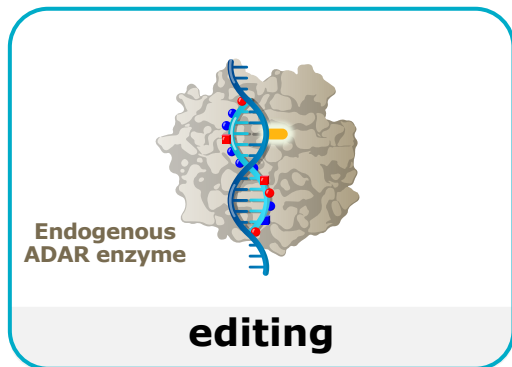


# Closing Remarks

Paul Bolno, MD, MBA  
*President and CEO*

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# Wave is building the leading RNA medicines company



Most versatile RNA medicines platform (PRISM™) in the industry



Best-in-class nucleic acid chemistry applicable across modalities

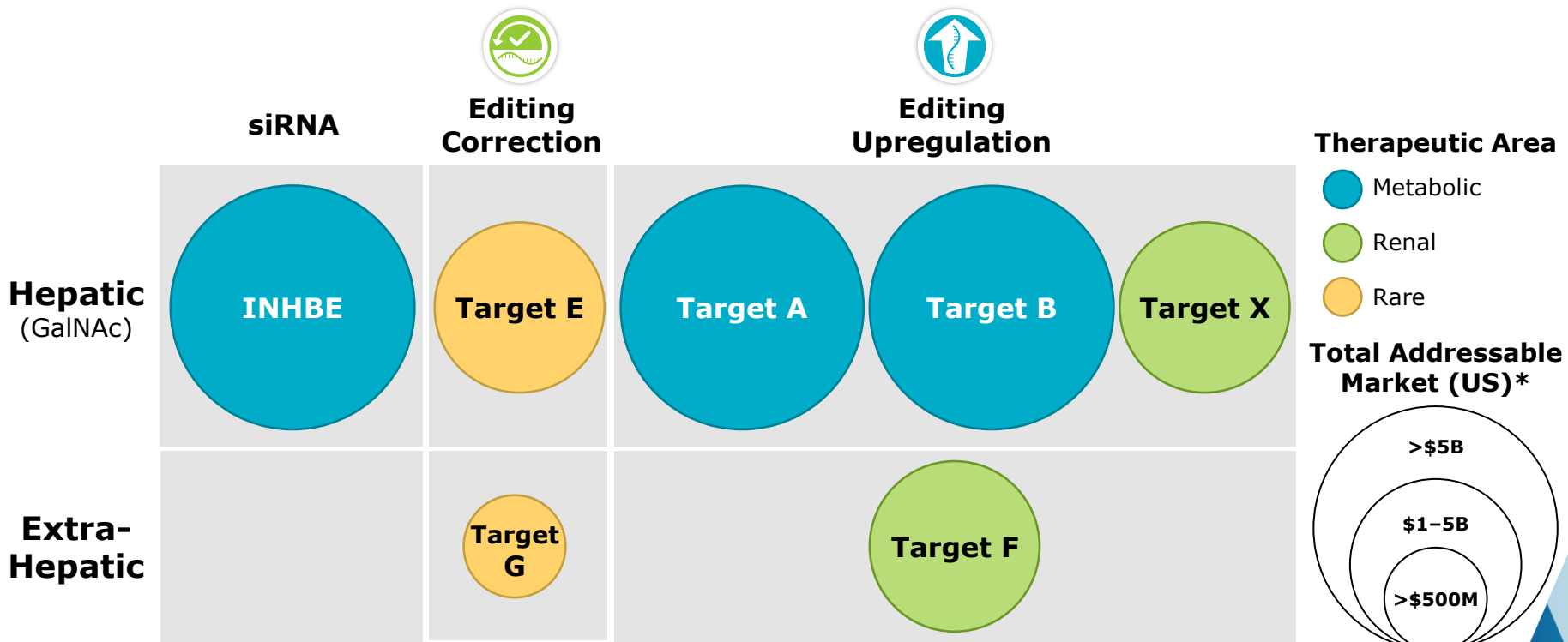


Ability to access novel / untapped areas of disease biology

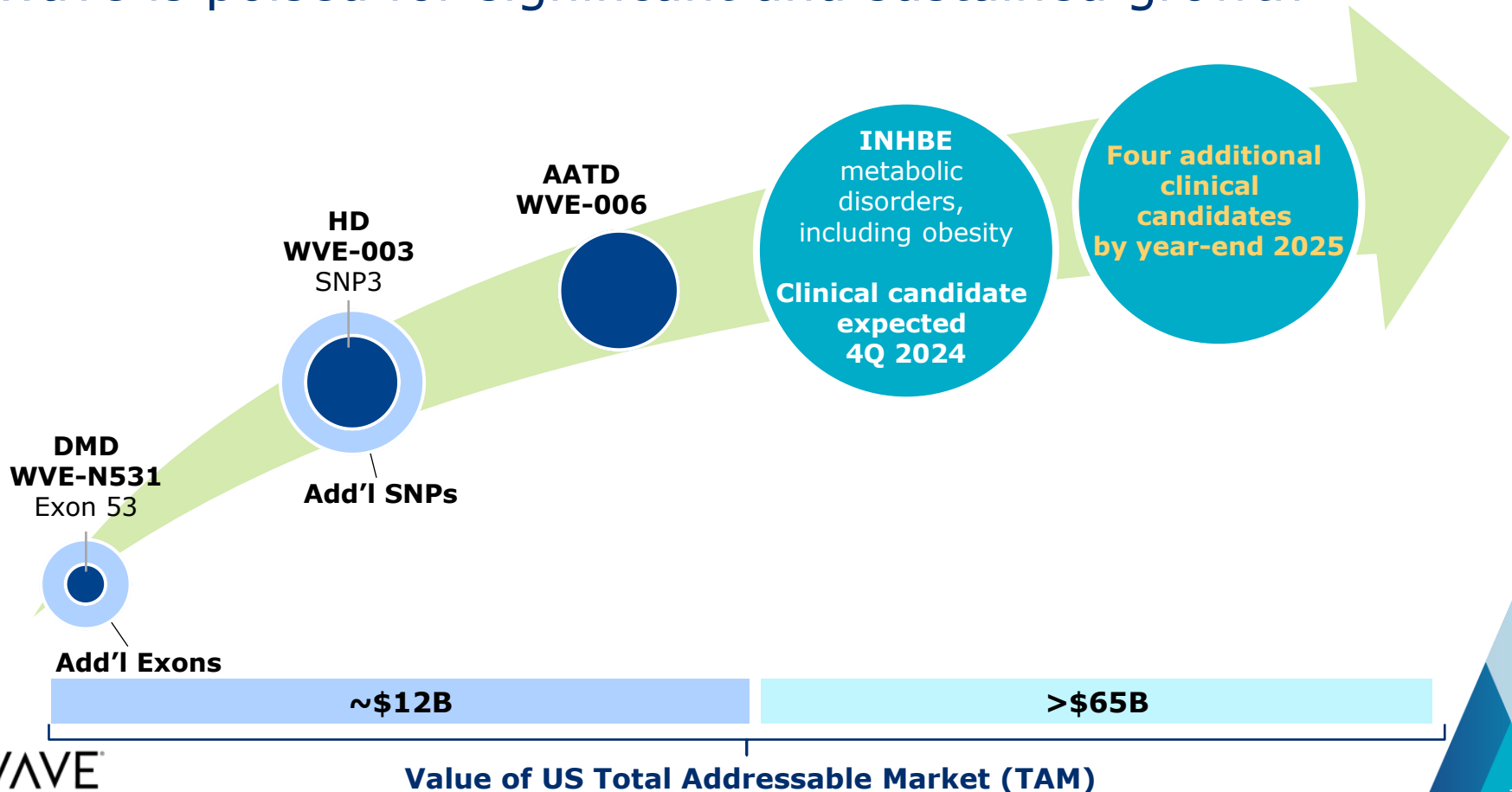


Platform learnings and clinical validation continue to increase probability of success

# Anticipate five new clinical candidates by year-end 2025



# Wave is poised for significant and sustained growth





**Paul Bolno, MD, MBA**  
*President and CEO*



**Anne-Marie Li-Kwai-Cheung**  
*Chief Development Officer*



**Chandra Vargeese, PhD**  
*Chief Technology Officer,  
Head of Platform Discovery Sciences*



**Kenneth Longo**  
*Vice President,  
Discovery Data Science*



**Ginnie Yang**  
*Senior Vice President,  
Translational Medicine*

# Q&A



# Thank you

For more information:  
[InvestorRelations@wavelifesci.com](mailto:InvestorRelations@wavelifesci.com)

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